

Micro-fluorimetry for the Biosciences

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BD Biosciences

Clontech
Discovery Labware
Immunocytometry Systems
Pharmingen



Need of Biology Research

Biological measurements

Ideally measure all chemicals in an organism with spatial and temporal resolution

- **Microscopy**

high spatial resolution to cell substructures, few cells, few parallel chemical measurements with subcellular resolution

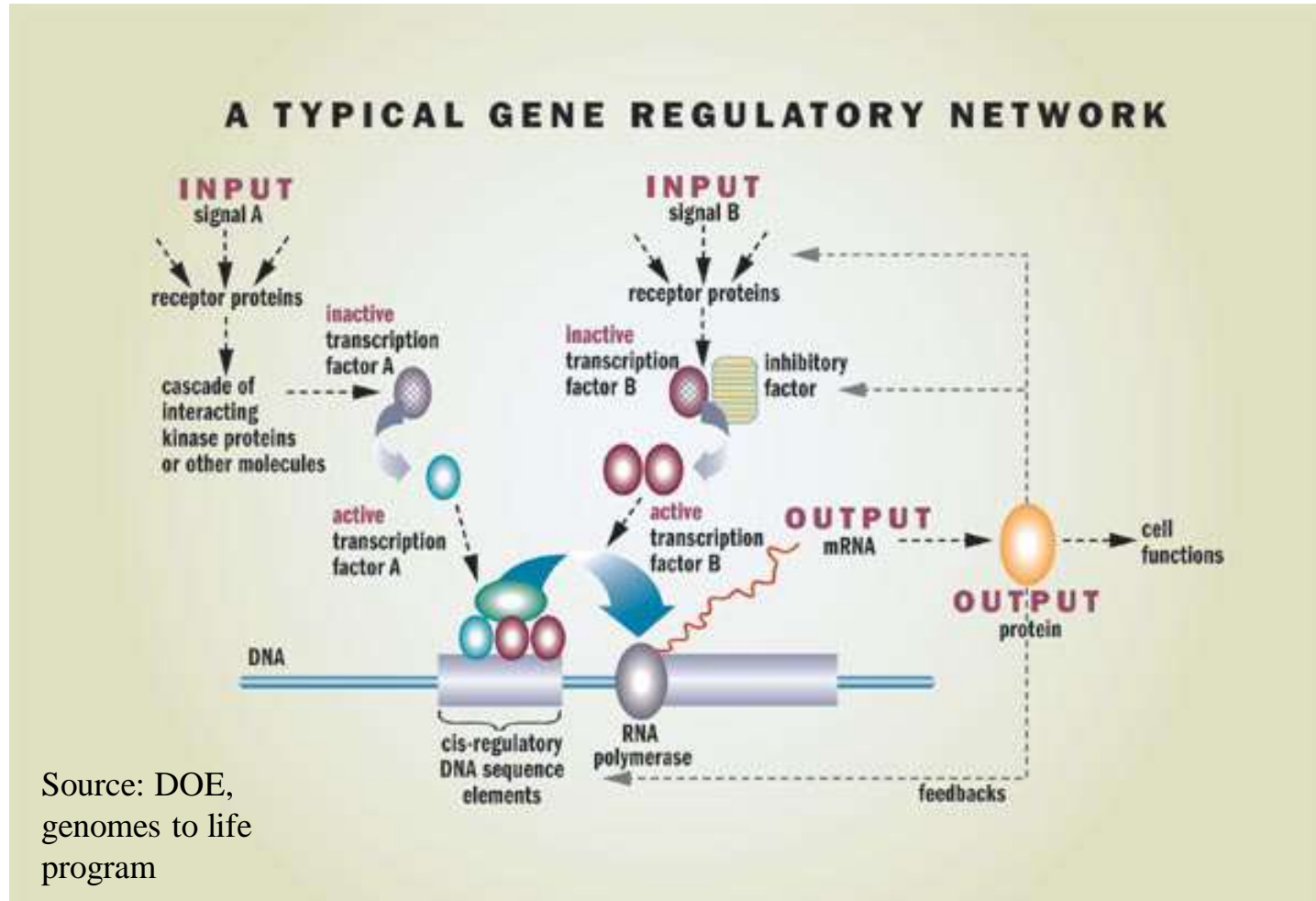
- **Flow cytometry**

spatial resolution to one cell, few to many cells, several parallel chemical measurements with one cell resolution

- **Arrays**

low spatial resolution to macroscopic cell ensemble level, many cells, many parallel chemical measurements cell sample resolution

Signal Transduction Network



System Components

Detection Instrument Considerations

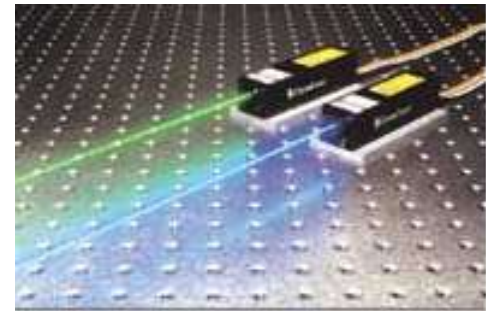
- **Light source** (wavelength, power, beam-shape)
- **Excitation optics** (direct beams vs. fiberoptics, multi-laser)
- **Emission optics** (collection efficiency NA, immersion optics)
- **Spectral filtering** (dispersive elements, filters, ...)
- **Detectors** (PMT, CCD, ...)
- **Signal processing** (analog vs. digital, data reduction)
- **System background** (electronic noise, photon statistics, ...)
- **Fluidics** (sheath flow vs undiluted sample, edge effects, focal depth)

Excitation light sources

High luminous density emitters

(490nm a desirable wavelength for fluorescein)

- Arc lamps (Hg-arc, ...)
- Gas lasers (Ar, Kr, ...)
- Solid state lasers

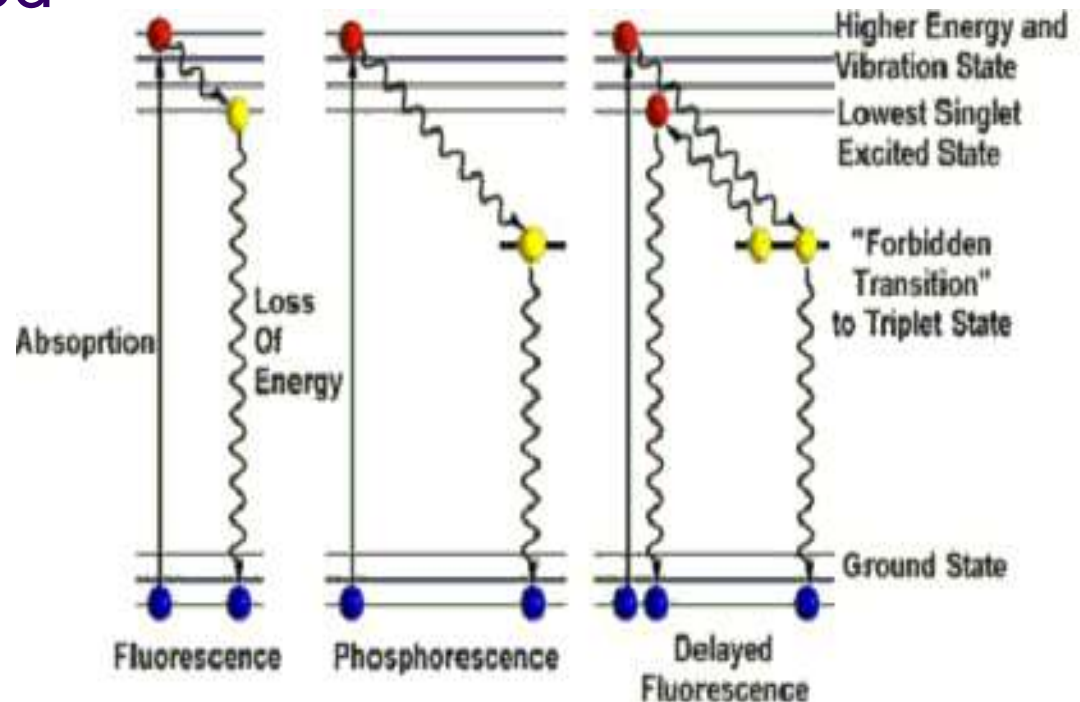


www.crystalaser.com

Fluorescence

- Absorbance
- Lifetime of excited state
- Stokes shift
- Quantum yield
- Saturation

- Intrinsic
- Extrinsic



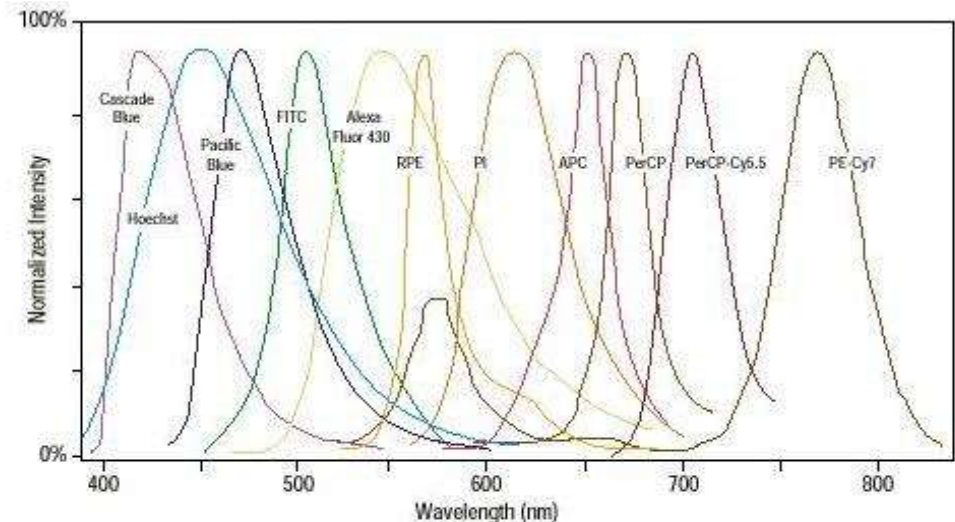
Source: www.olympusmicro.com

Fluorescent Dyes

- Unsaturated organic molecules (photobleaching)
- Energy-transfer dyes
- Lanthanide derived pigments
- Quantum dots

Dye combinations for multi-color fluorescence

- Single vs. multi-laser excitation
- Spectral overlap

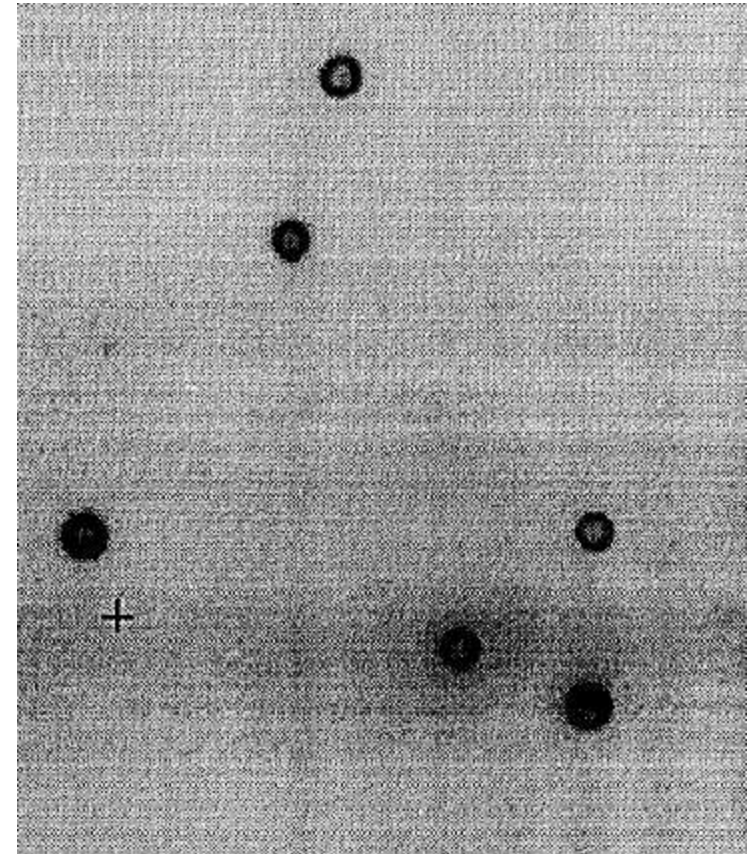
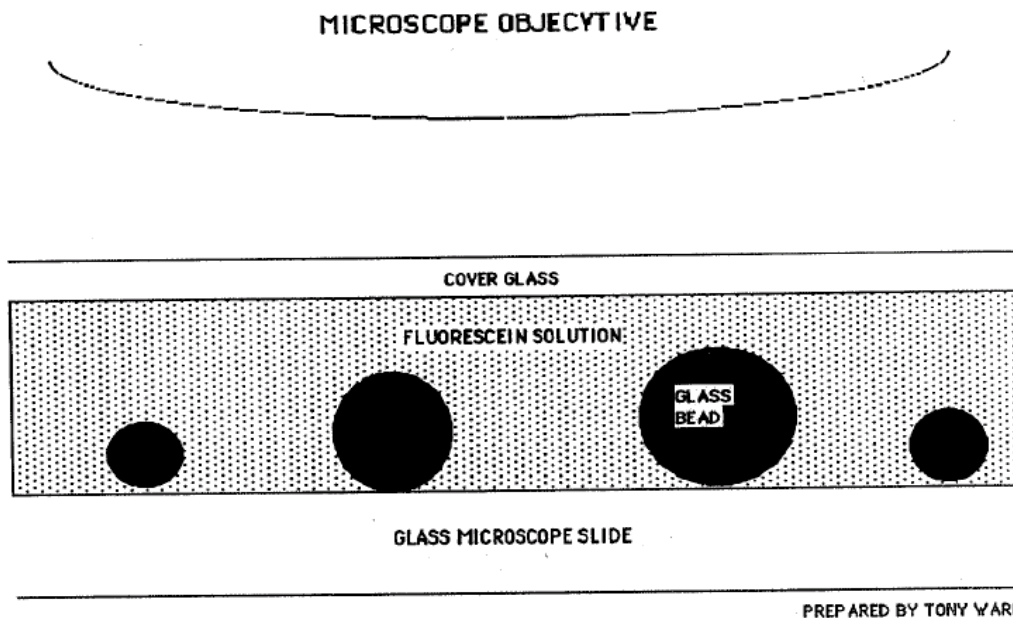


Dye Spectra From the FACSaria User Guide

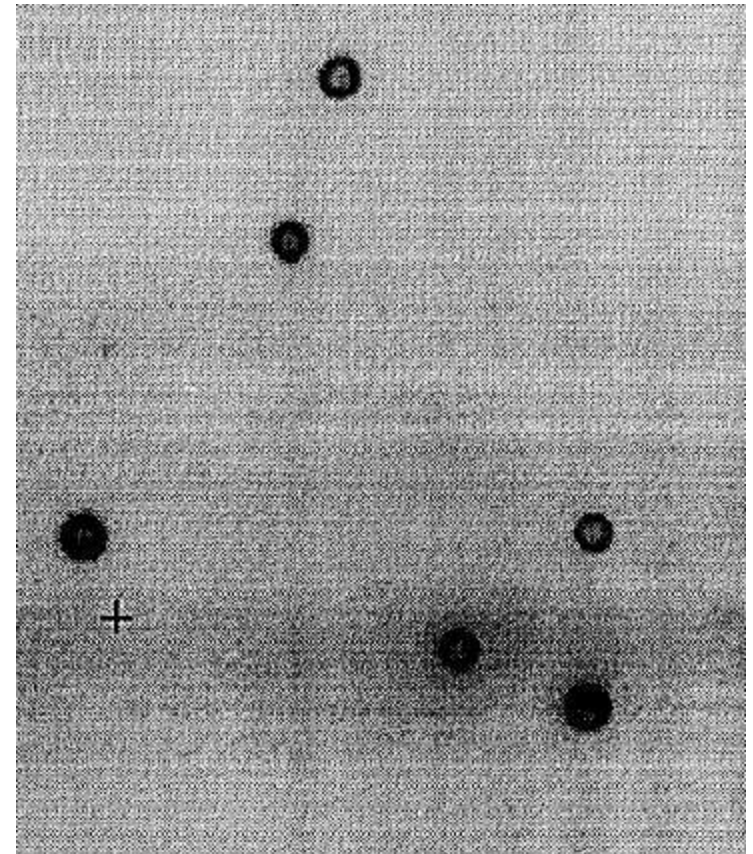
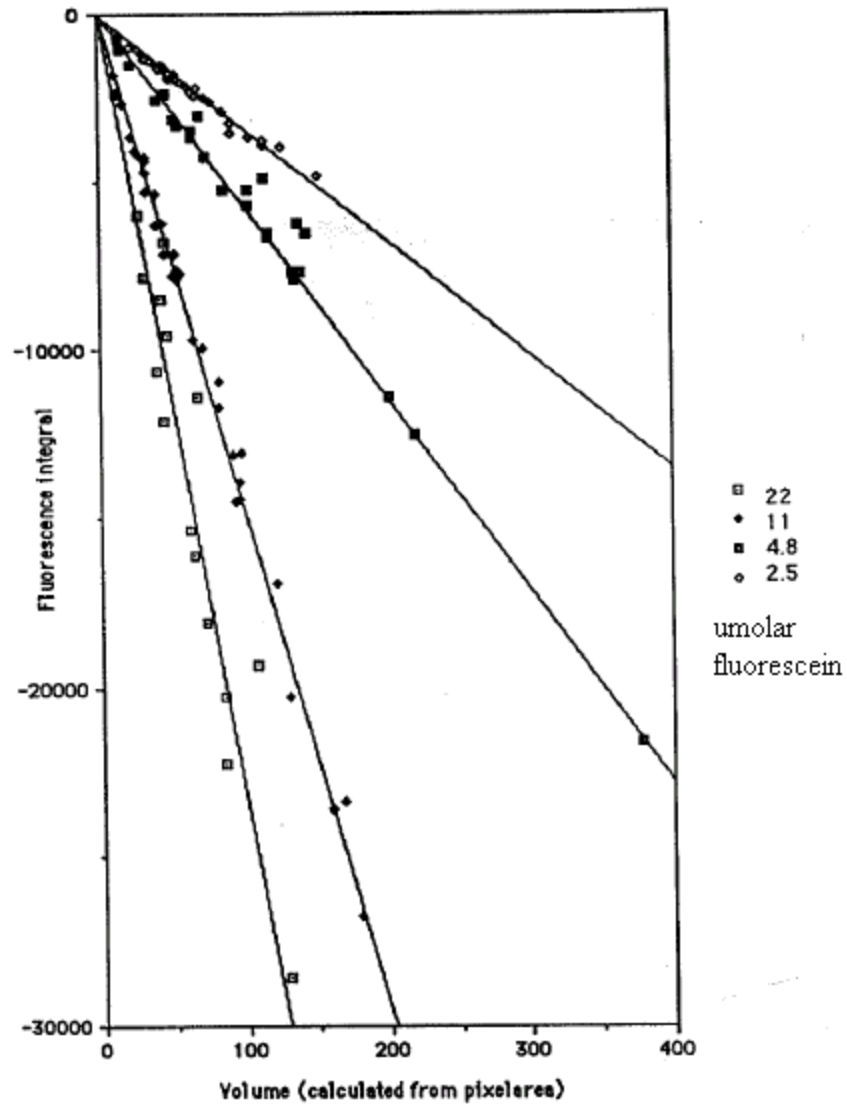
Fluorescence Measurements of Chemical Properties

- Intensity Concentration
- Lifetime Background reduction
- FRET (energy transfer) Proximity
- Polarization Rotational mobility
- Fluorescence correlation Lateral mobility

Fluorescence Quantitation by Volume Exclusion (1)



Fluorescence Quantitation by Volume Exclusion (2)

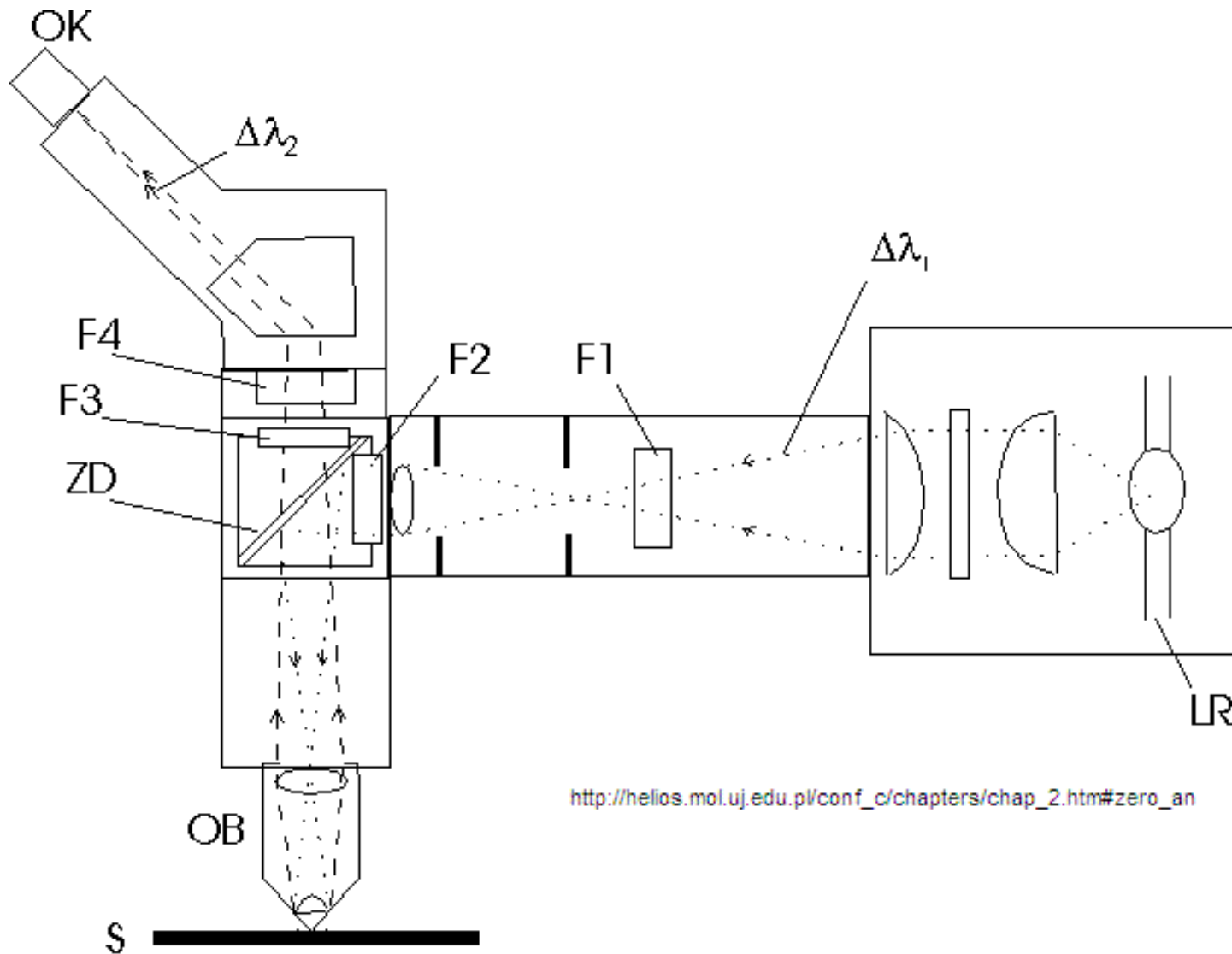


Assay considerations

- Reagent selection
- Selection of labels (spectral properties, overlap)
- Homogeneous vs. non-homogeneous
- Intrinsic fluorescence vs. use of labels
- Specimen and reagent auto-fluorescence
- Reference samples

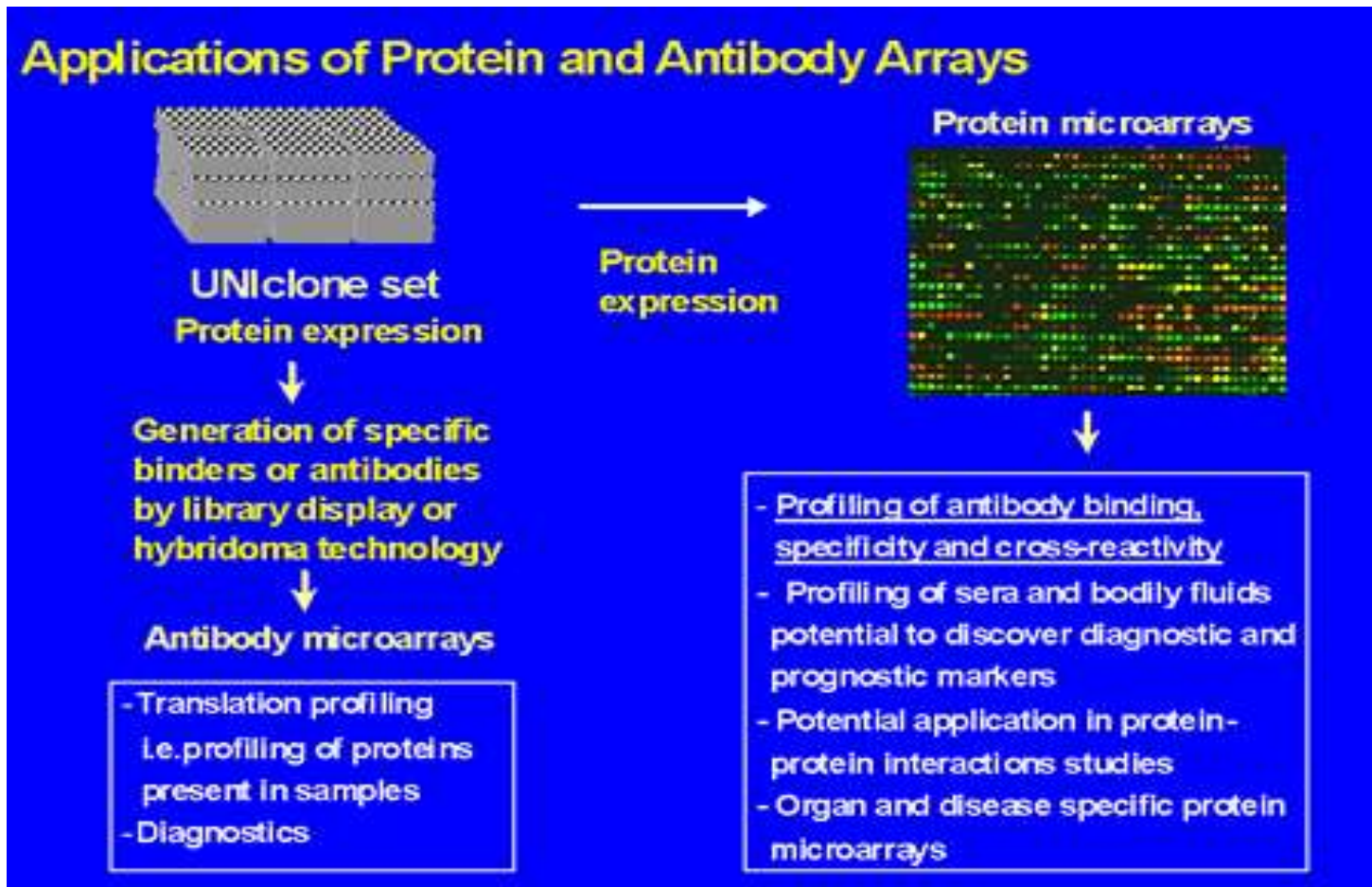
Systems, which use fluorescence

Fluorescence Microscope



http://helios.mol.uj.edu.pl/conf_c/chapters/chap_2.htm#zero_an

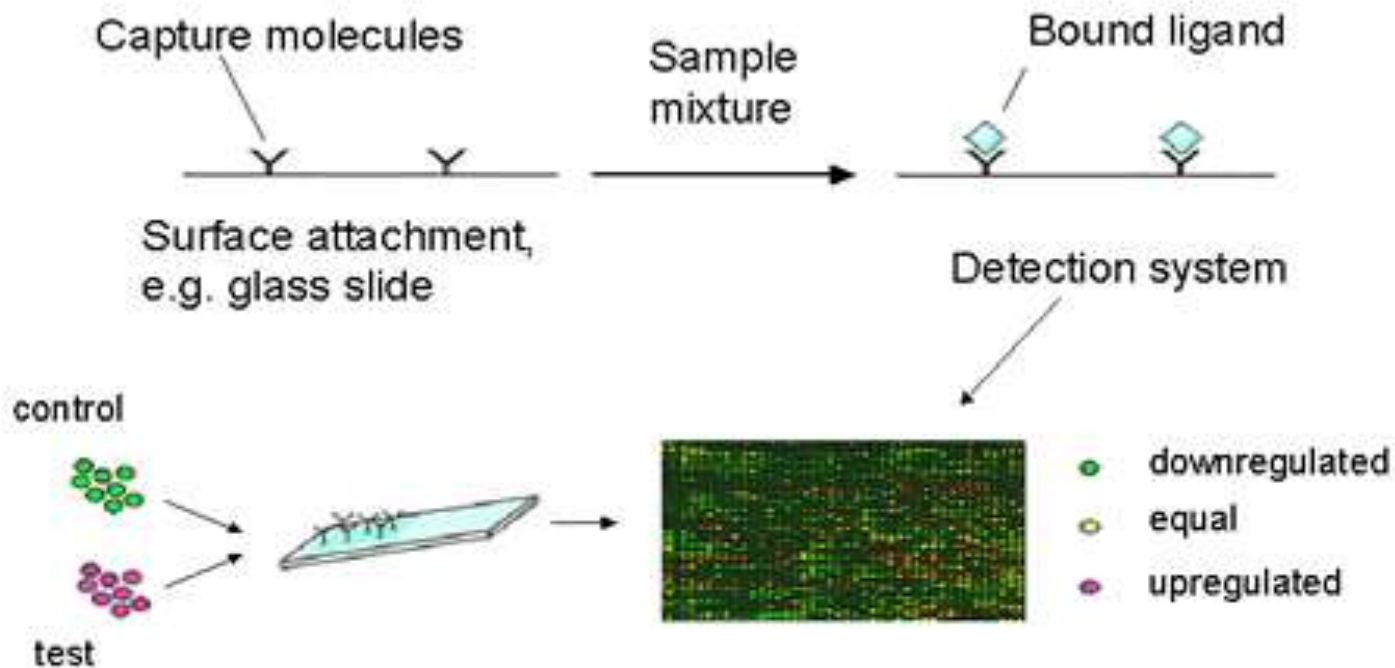
Micro-arrays



Dolores Cahill, Max-Planck-Institute of Molecular Genetics

Micro-arrays

Antibody microarray principle



e.g. to compare protein expression, mix Cy3/Cy5 labelled samples

http://www.functionalgenomics.org.uk/sections/resources/protein_arrays.htm#related

Micro-arrays

Antibody arrays: Detection strategies

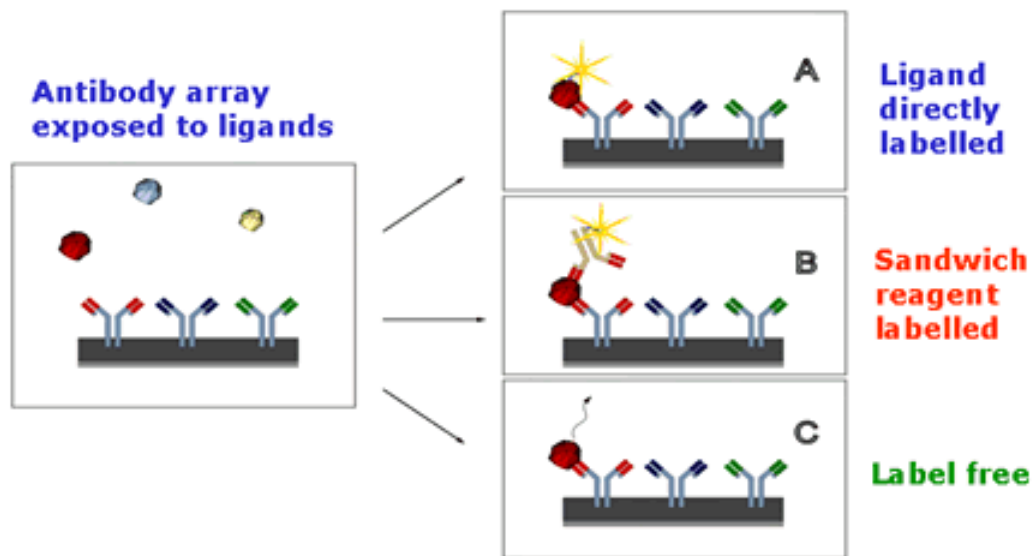
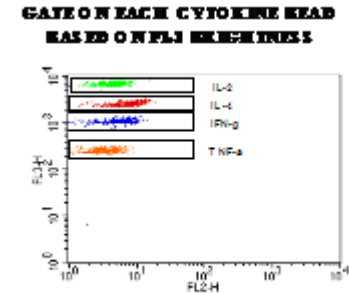
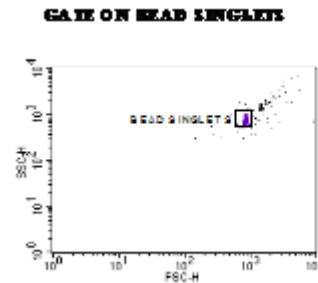
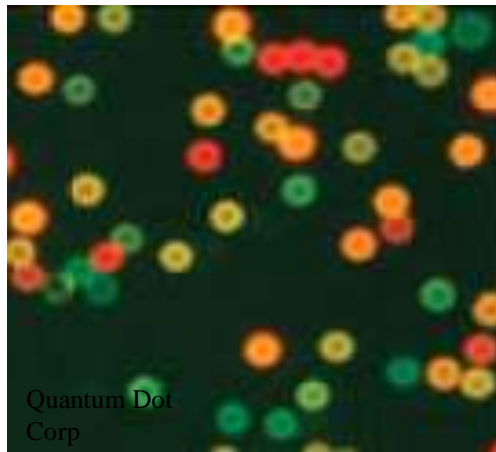


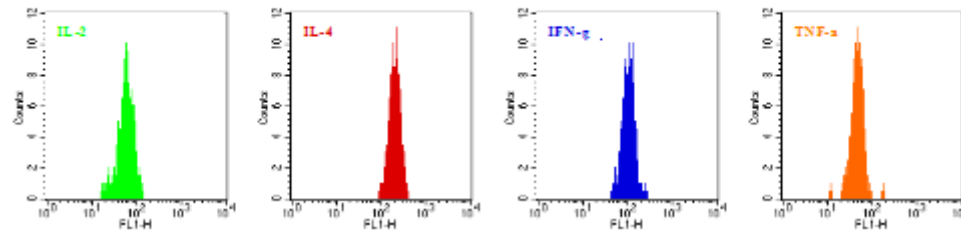
Figure prepared by
Jonas Jarvius,
Uppsala (from Taussig
and Landegren,
Targets 2:169-176,
2003)

http://www.functionalgenomics.org.uk/sections/resources/protein_arrays.htm#detection

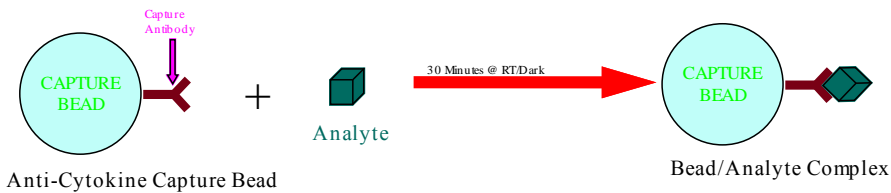
Cytometric Bead Array Assays



ANALYZE FOR PRESENCE OF EACH CYTOKINE BASED ON FLUORESCENCE YIELDING ON FL1



Step 1: Analyte Capture



Step 2: Detector Antibody Binding

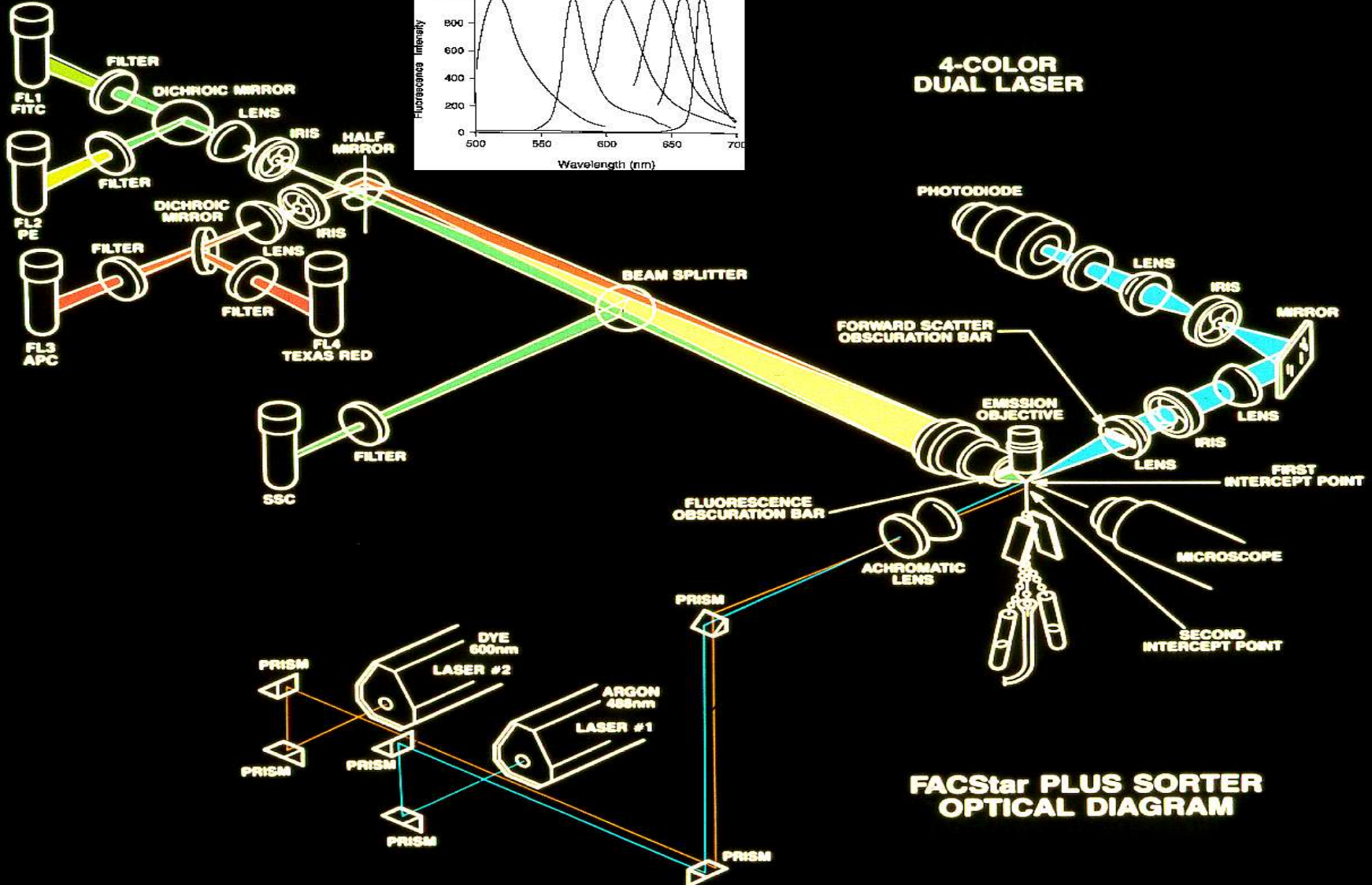
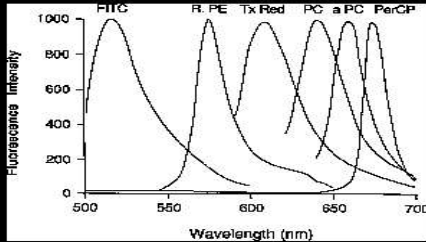


Step 3: Reporter Molecule Binding



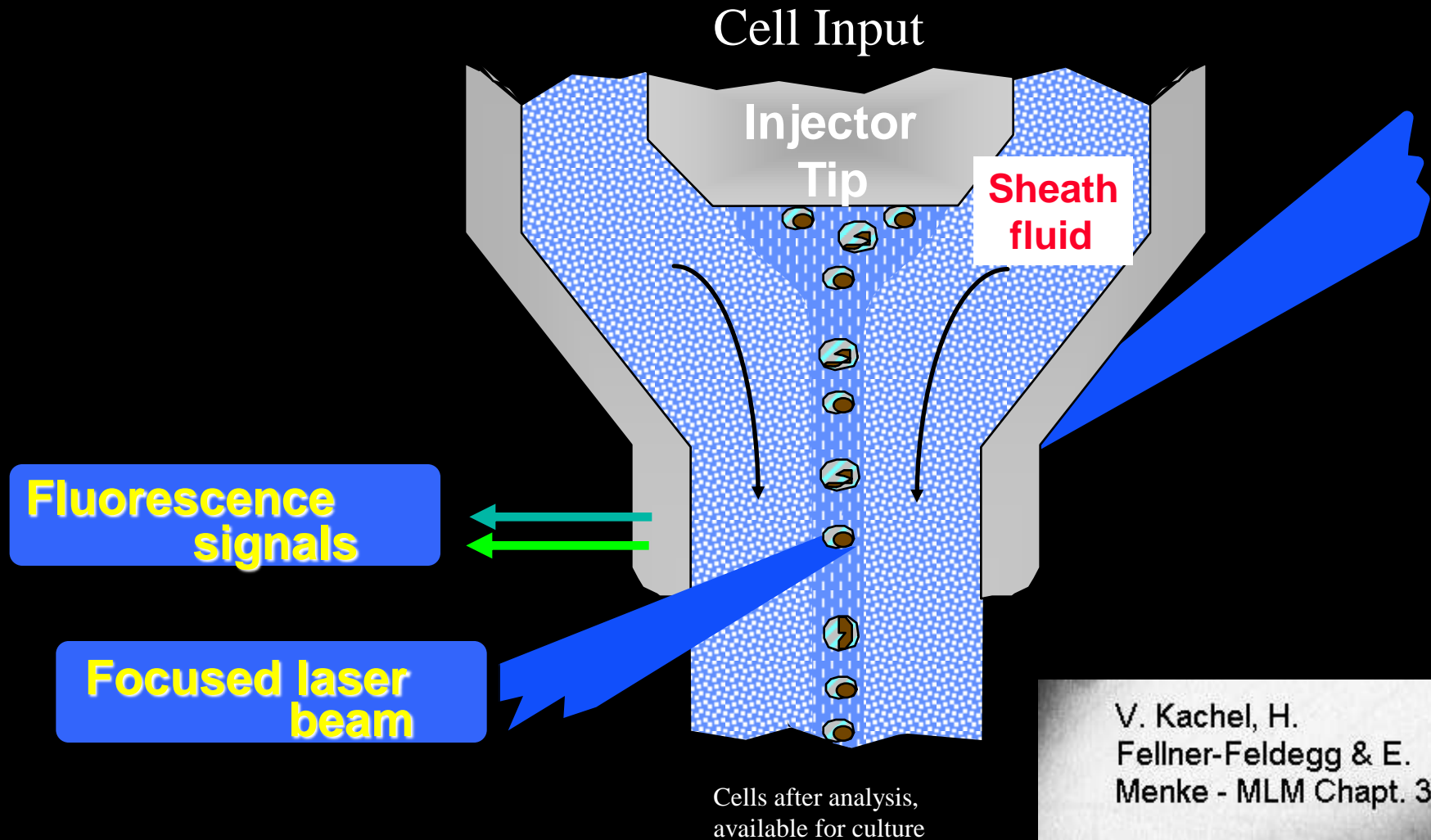
“Bead Array” assays

Flow Cytometers



FACStar PLUS SORTER OPTICAL DIAGRAM

Flow Cytometer Fluidics



Applications in research

Applications of Microfluorimetry

Measurements

- specific structures by immunofluorescence or in-situ hybridization
- protein content
- DNA and RNA with fluorogenic dyes
- gene expression with fluorescent proteins
- auto-fluorescent components
- enzyme activity with fluorogenic substrates
- pH and other cations with ion-specific probes
- redox potential
- ...

Applications of Microfluorimetry

- Cell sorting
- Multi-parameter cell subset analysis
- Organelle visualization
- Protein translocation
- Large scale cell composition changes
- Single fluorescent molecule detection
(PCR vs. direct)
 - Nucleic acid fragment sizing
 - Protein-ligand interactions
 - Virus counting

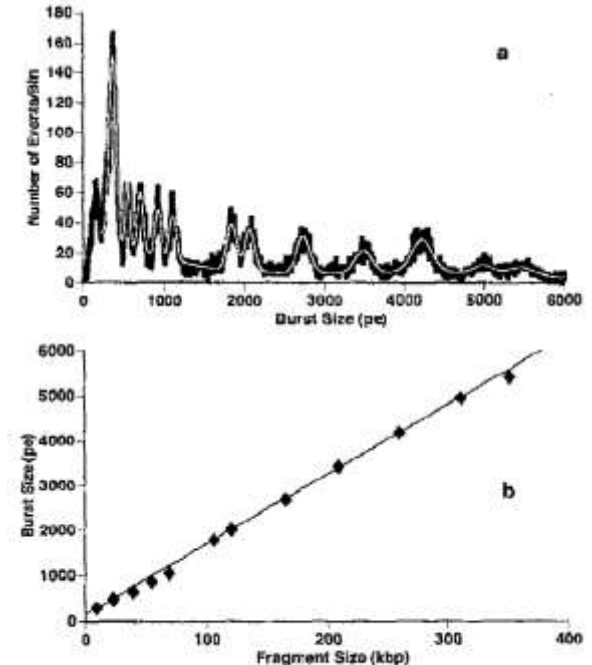
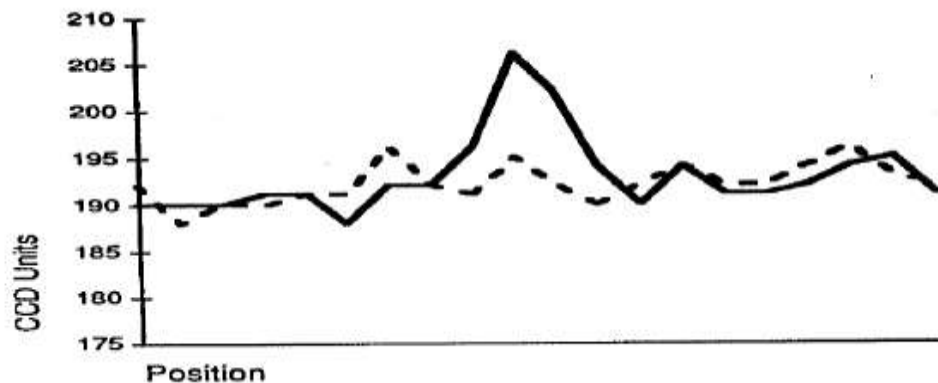
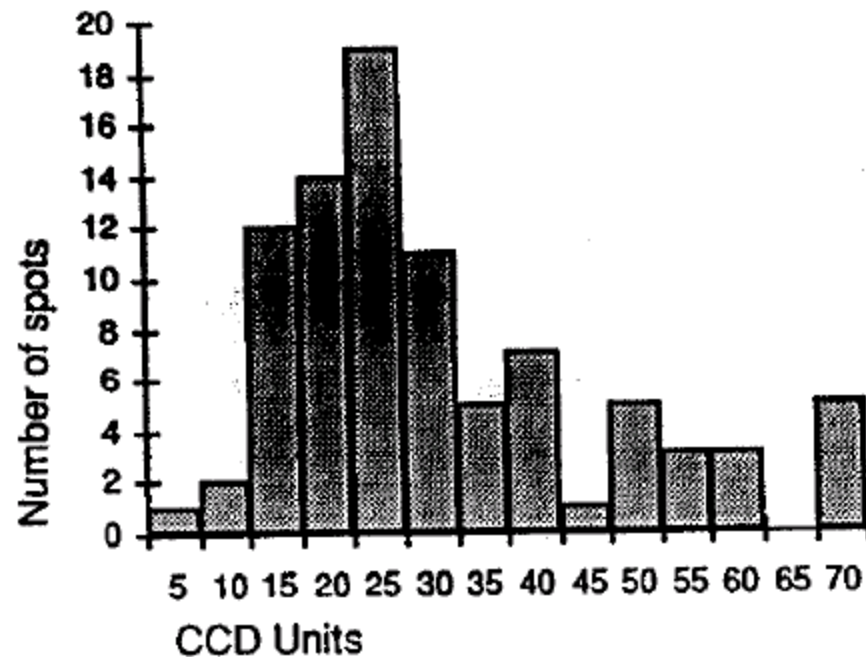
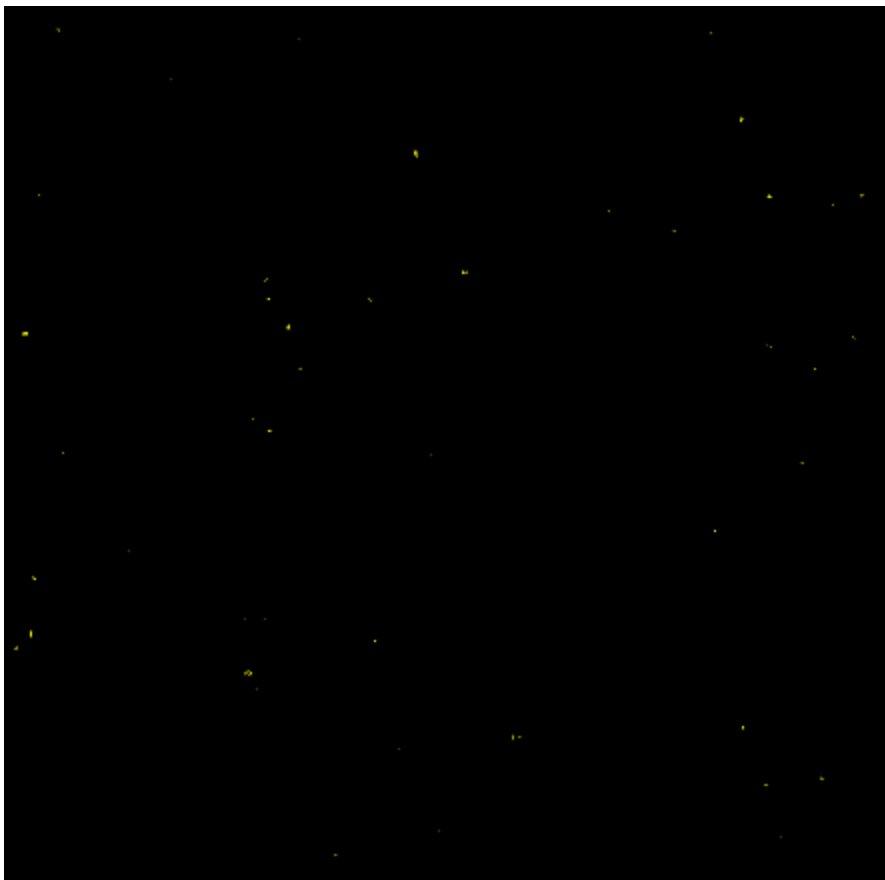


Fig. 2. a) Histogram of the fluorescence burst sizes of *SmaI* digestion of *S. aureus* DNA. Bin width was 10 pe.

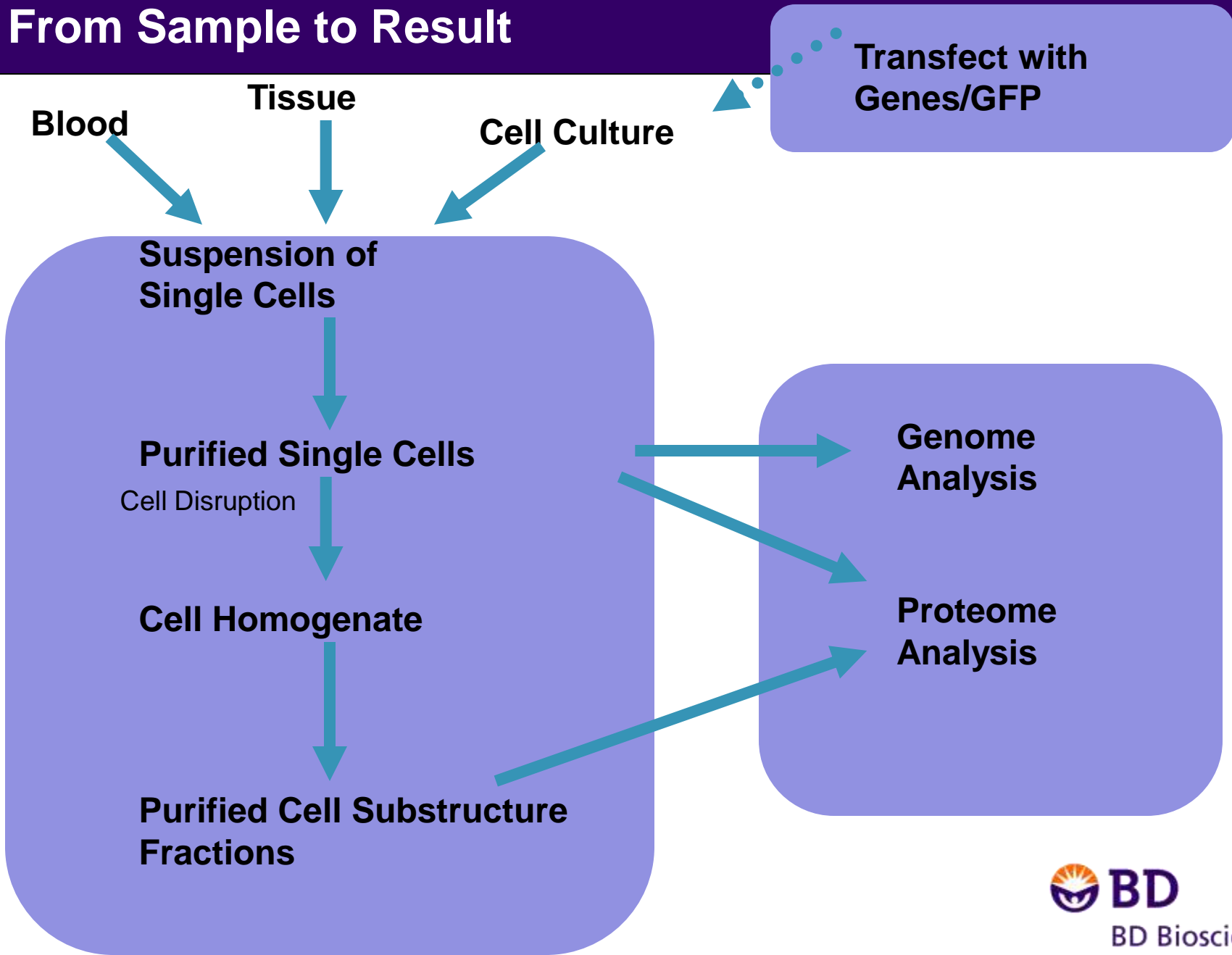
From: Huang Z et al. in

Cytometry 35: 169-175 (1999)

Single Molecule Detection



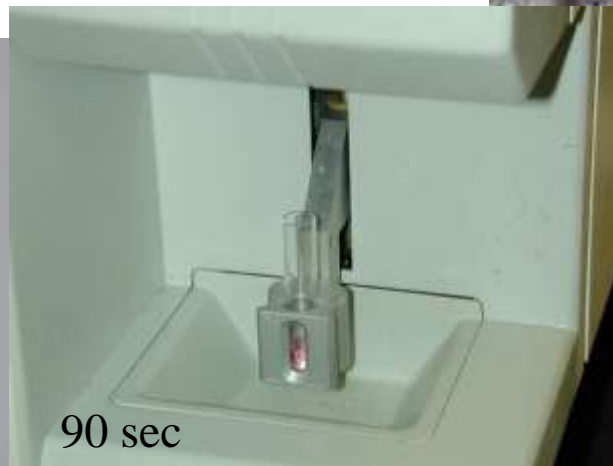
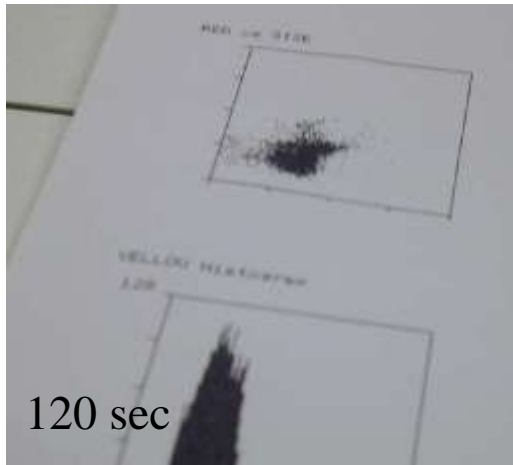
Cellular Proteomics: From Sample to Result



Applications in Bio-defense

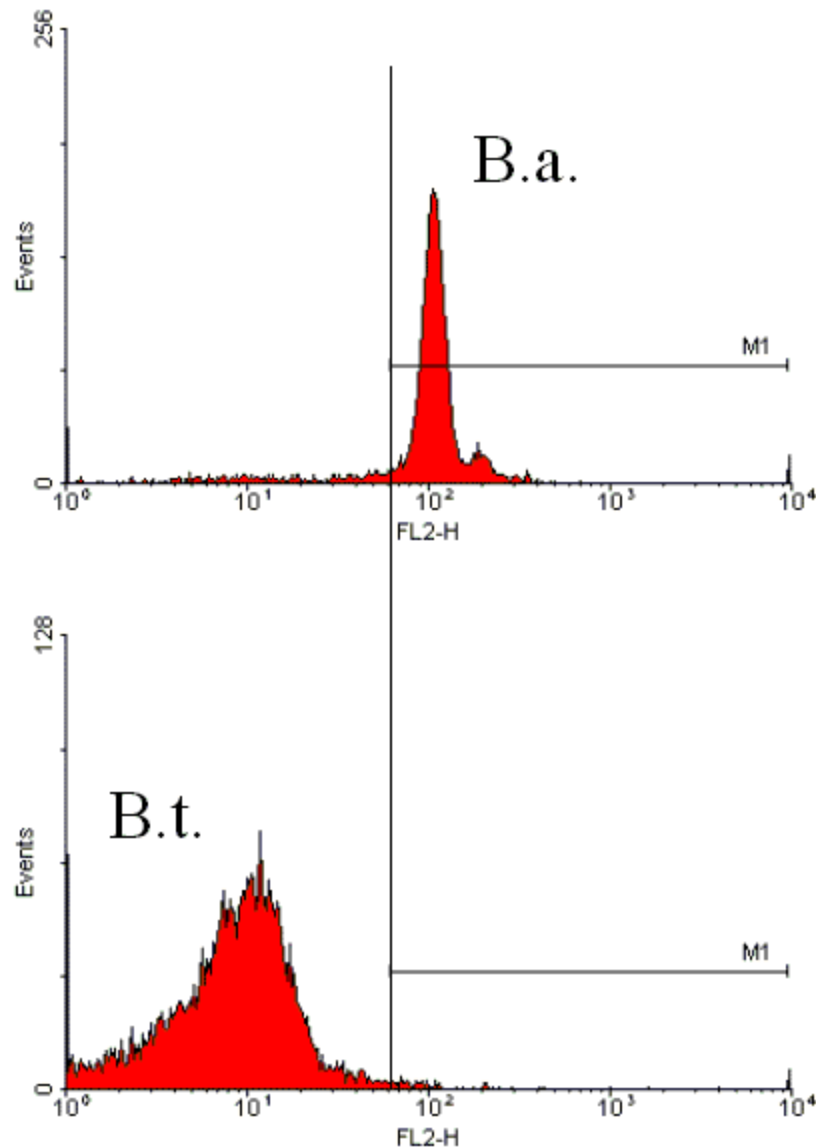
Flow Cytometry for Microbiology

Rapid TVC Assay



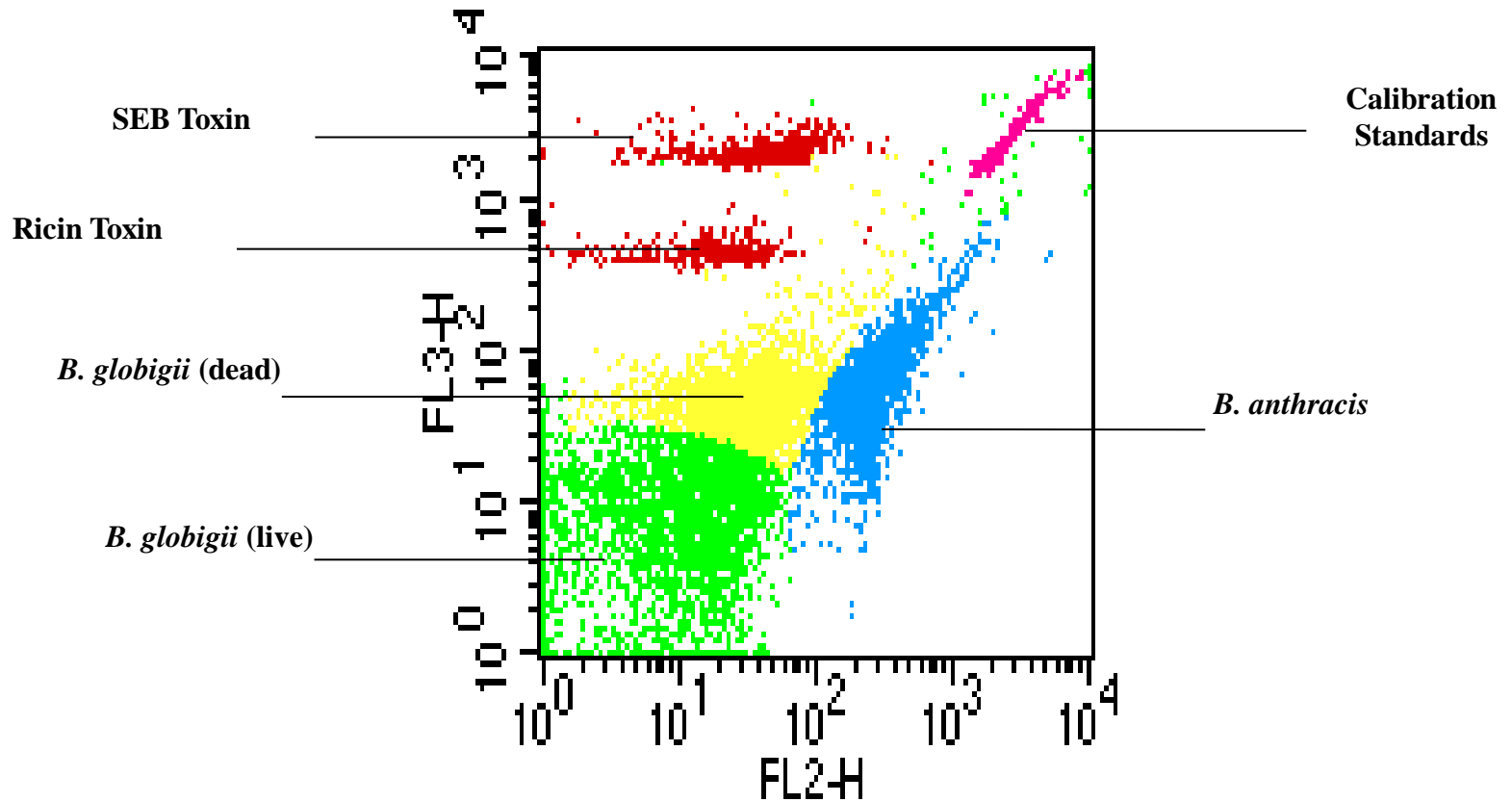
Rana Alsharif, BD Biosciences

Rapid Bioagent Identification



Fluorescence based detection and identification of *Bacillus anthracis* within 10 minutes after receiving a sample using widely available instrumentation

Simultaneous Measurements of Multiple Agents



Applications in Clinical Diagnostics and Monitoring

Clinical Tests (CD34 counting)

- Large scale protein analysis in patient serum with arrays
- Cell subset analysis (special hematology)
 - CD4
 - CD34
 - L&L
- Histopathology

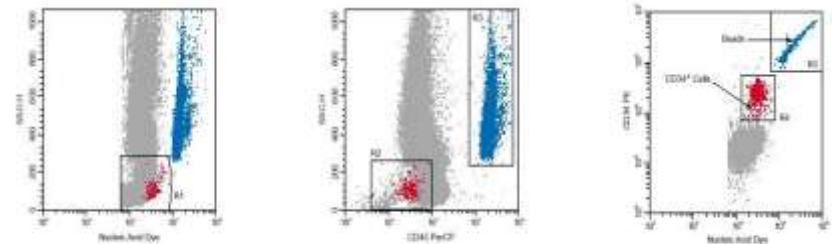
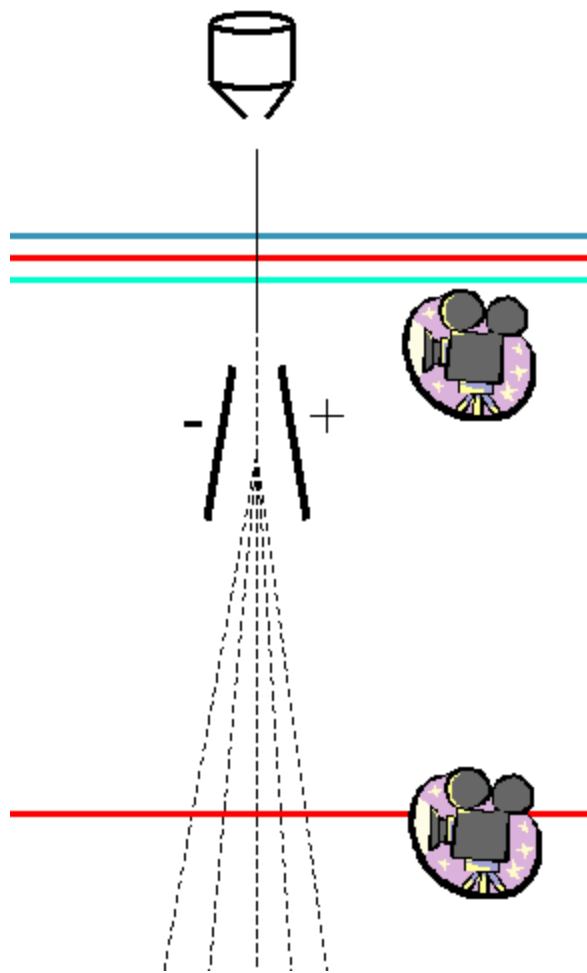


Figure 3. ProCOUNT Multiparameter Data Analysis Gating Strategy

Therapy

Cell Sorting



CD34 progenitor cell sorting

Isolate human blood progenitor cells for cell transplantation

- CD34 at about 1% in mobilized peripheral blood
- for high purity several parameters are used
- $2-5 \times 10^6$ CD34 cells needed for treatment
- analysis rate maximally at 10^5 cells sec^{-1}
- sort rate at 2×10^4 cells sec^{-1}
- several hours of sorting required

Outlook

Challenges

- Better use of single molecule sensitivity
- Absolute quantitation
- Better multiplexing
- Efficient use of intrinsic fluorescence

END

BACKUP SLIDES

Limits of Detection by CCD Microscopy

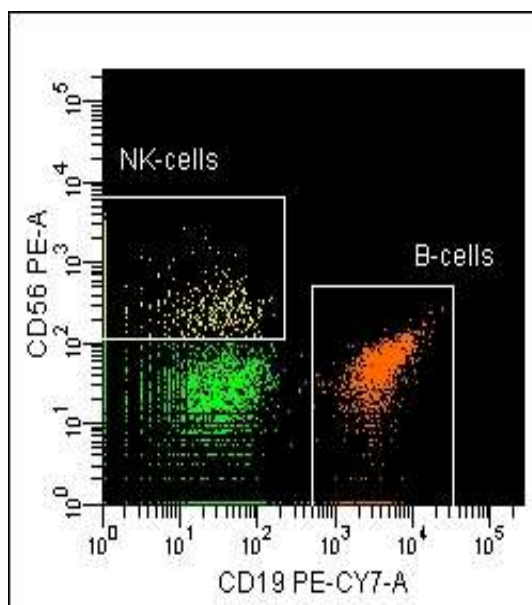
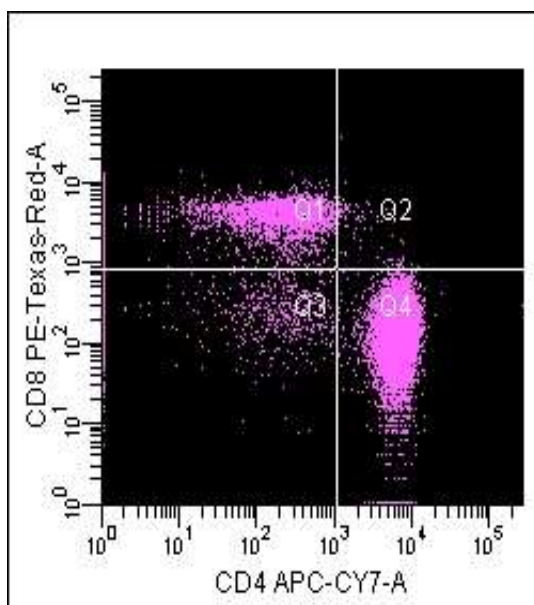
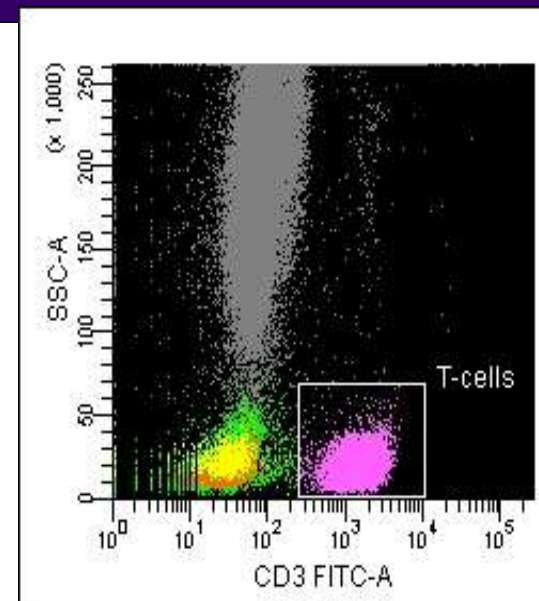
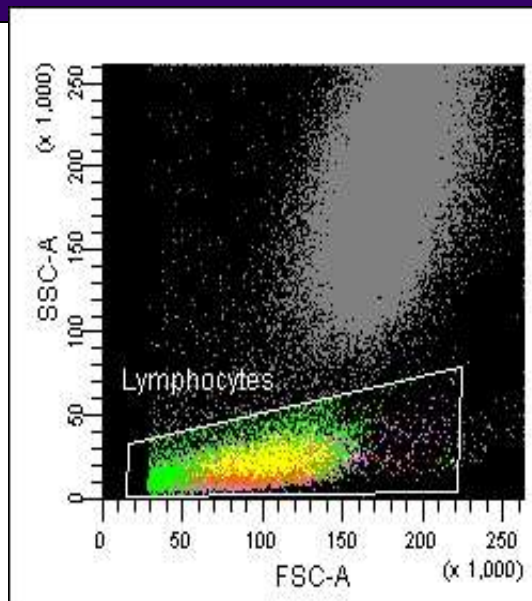
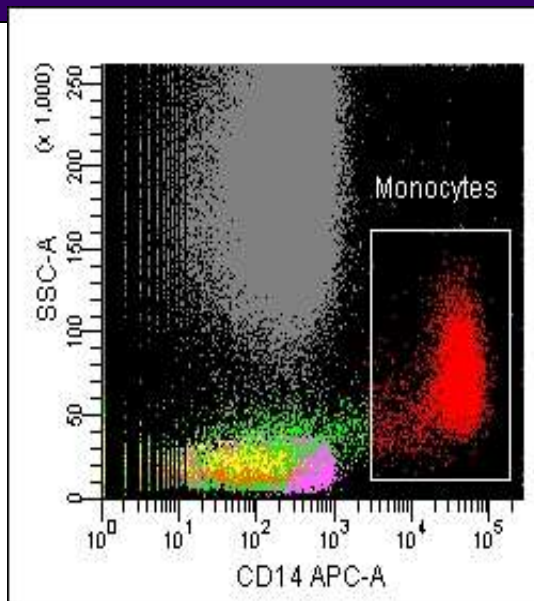
Diether Recktenwald, Janette Phi-Wilson, Ben Verwer 1993

Fluorescein	1000 Molecules/Pixel
R.-Phycoerythrin	15 Molecules/Pixel

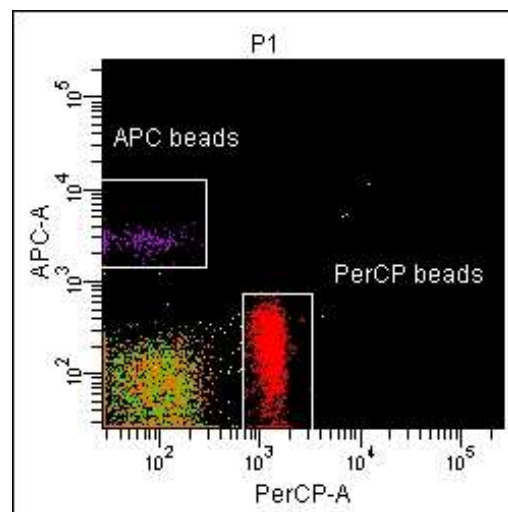
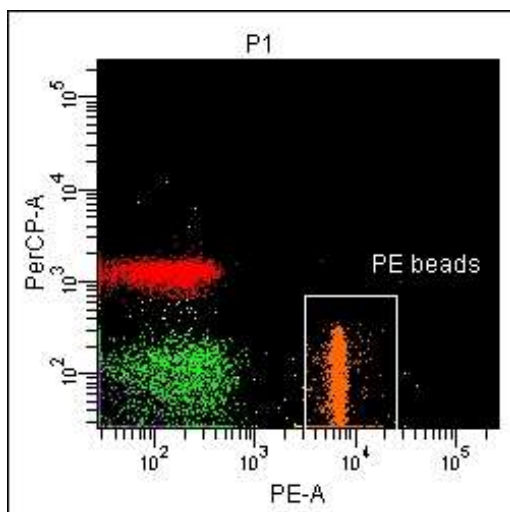
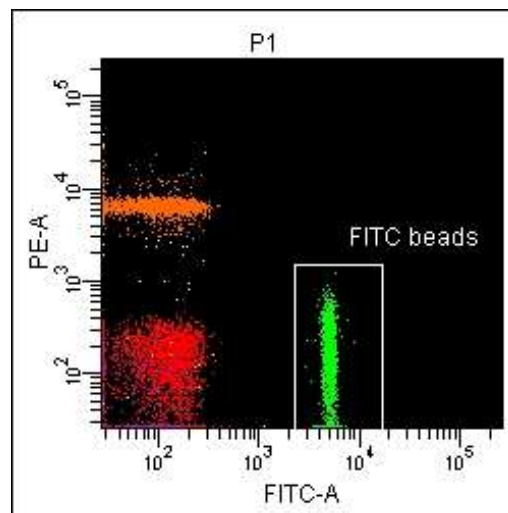
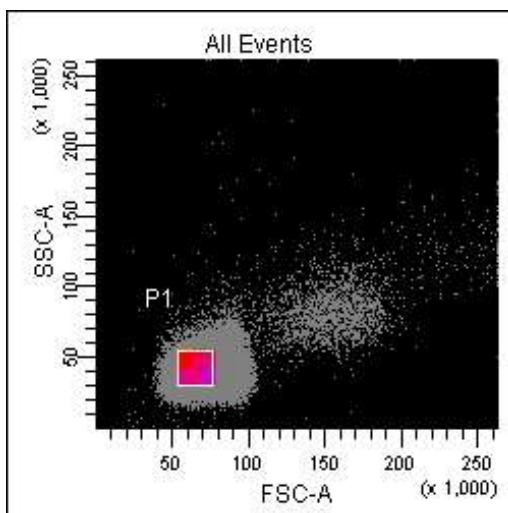
Factors limiting practical use:

- biological autofluorescence
- photo-bleaching
- diffusion

Multi-color analysis (6-color example)

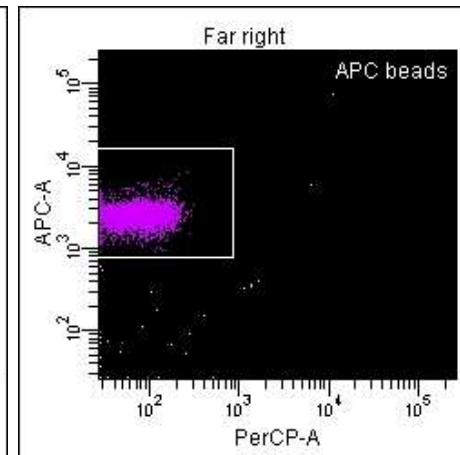
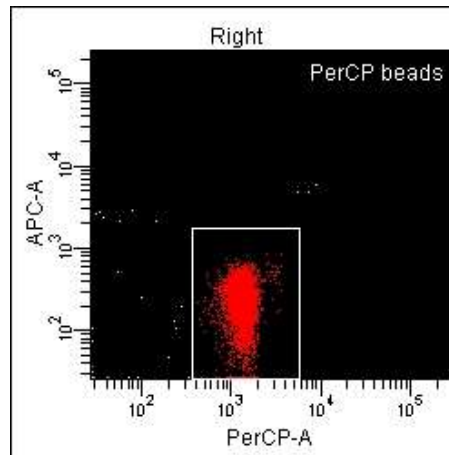
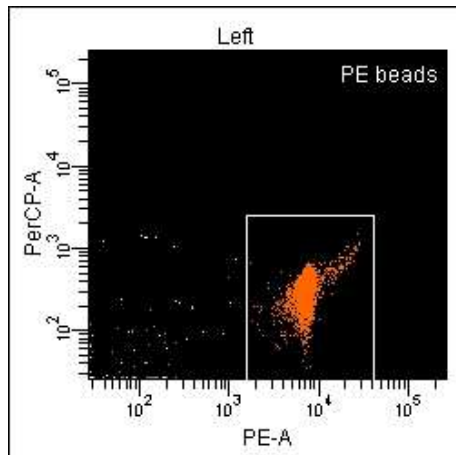
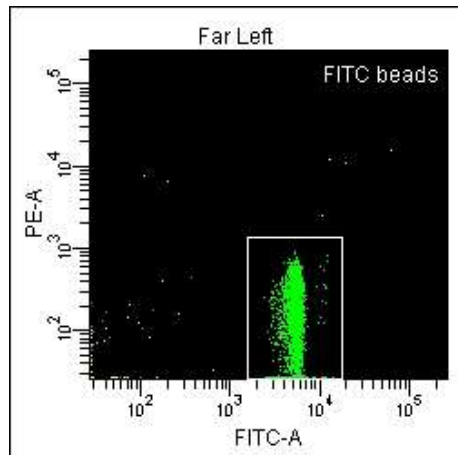


Digital Cytometry FACS (Pre- Sort)



45 psi
64 kHz
20,000 events/s

Digital Cytometry FACS (Purity)



FITC

8.2%

99.1%

PE

9.5%

99.2%

PerCP

8.0%

99.5%

APC

0.6%

99.4%

Abstract

- The understanding of complex biological systems requires the quantitative measurement of DNA, RNA, proteins, and metabolites in-vivo with high sensitivity. Micro-fluorimetry with modern light sources and detectors has demonstrated single molecule sensitivity. Instrumentation and assays with these capabilities for the measurement of cellular properties and secreted proteins will be discussed, including diagnostic and therapeutic uses of the technology.
-
- Suggested reading:
- 1.Mandy F, Varro R, Recktenwald D (2003) Flow Cytometry Principles in Biomedical Photonics Handbook, Tuan Vo-Dinh ed, CRC Press 2003
- 2.Enderlein J., Ambrose WP, Goodwin PM, Keller R (1999) Fluorescence Detection of Single Molecules, <http://www.joerg-enderlein.de/pubs/book99.pdf>
- 3.Baumgarth M, Roederer M (2000), JIM 243, 77. A practical approach to multicolor flow cytometry for immunophenotyping (<http://facs.scripps.edu/BRJIM.pdf>)