



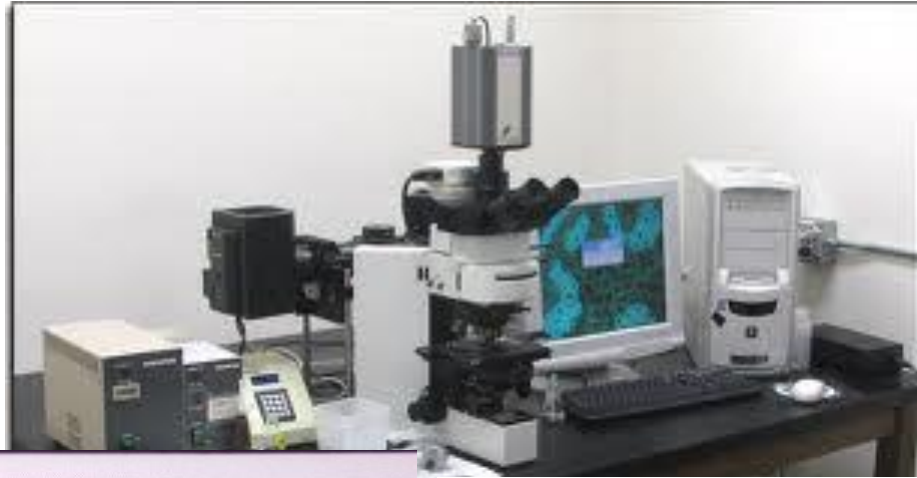
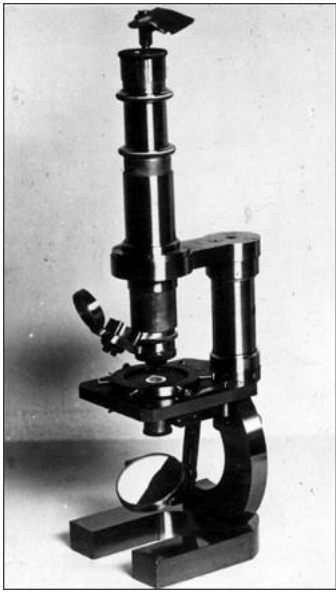
# State of the Art Cytometry & Emerging Technologies for Single Cell Analysis

Diether Recktenwald, BD Biosciences, retired  
Desatoya LLC, Reno NV, USA  
Email: [diether@desatoya.com](mailto:diether@desatoya.com)  
<http://www.desatoya.com>

# Biology Research Targets and Tools

<b>Organism</b>	NMR	Contrast agents
	X-ray imaging	Affinity reagents
<b>Organ</b>	Ultrasound	- antibodies
	2-photon imaging	- probes
<b>Tissue</b>	In-vivo cytometry	Enzyme substrates
	Light microscopy	Labels
<b>Single Cell</b>	Electron microscopy	- absorbance
	Flow cytometry	- fluorescence
<b>Organelle</b>	Cell imaging	- element tags
	NA sequencing	
<b>Macromolecule</b>	Mass spectrometry	
	TIRF microscopy	
<b>Small molecules</b>	Electrophoresis	Sample prep

# Early & Modern Flow Cytometry and Imaging Microscopy



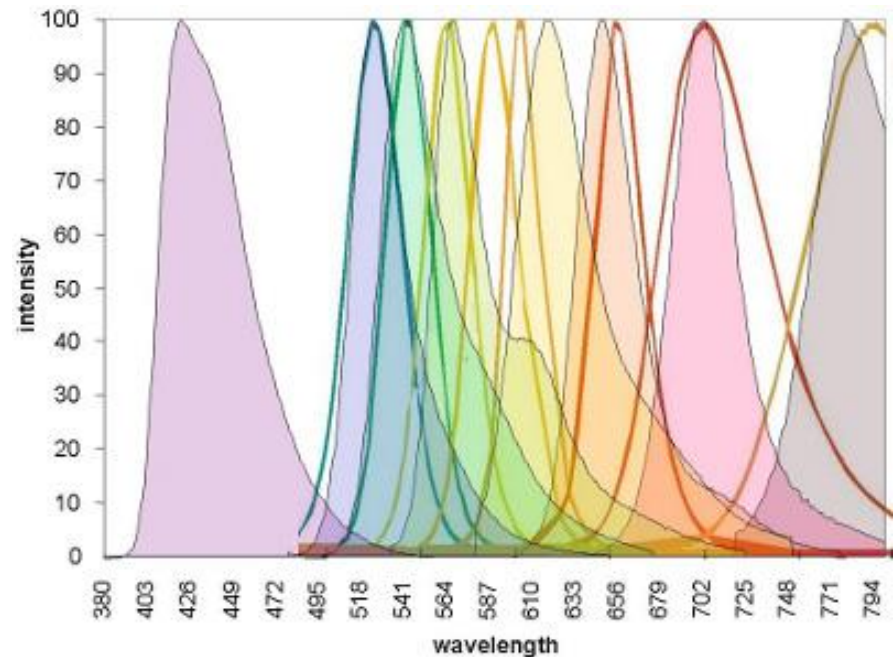
# Flow and Imaging Cytometry Features

Single particle (cell) analysis with

- High sensitivity (single molecule sensitivity by fluorescence) I,F
- Wide dynamic count range ( $10^3$  to  $10^7$  cells  $\text{mL}^{-1}$ ) F
- Particle sizes from 0.2 to 20  $\mu\text{m}$  F, I
- High analysis rates to  $\sim 10^5$  particles  $\text{sec}^{-1}$  F
- Direct size and 3D spatial information I
- Multi-color fluorescence, multi-parameter analysis F,I
- Wide dynamic range for fluorescence ( $10^5$ ) F
- Direct kinetic measurements I
- Viable cells can be re-covered F,(I)
- Measurement of adherent cells I
- Good ease-of-use F,(I)

# Physical parameters

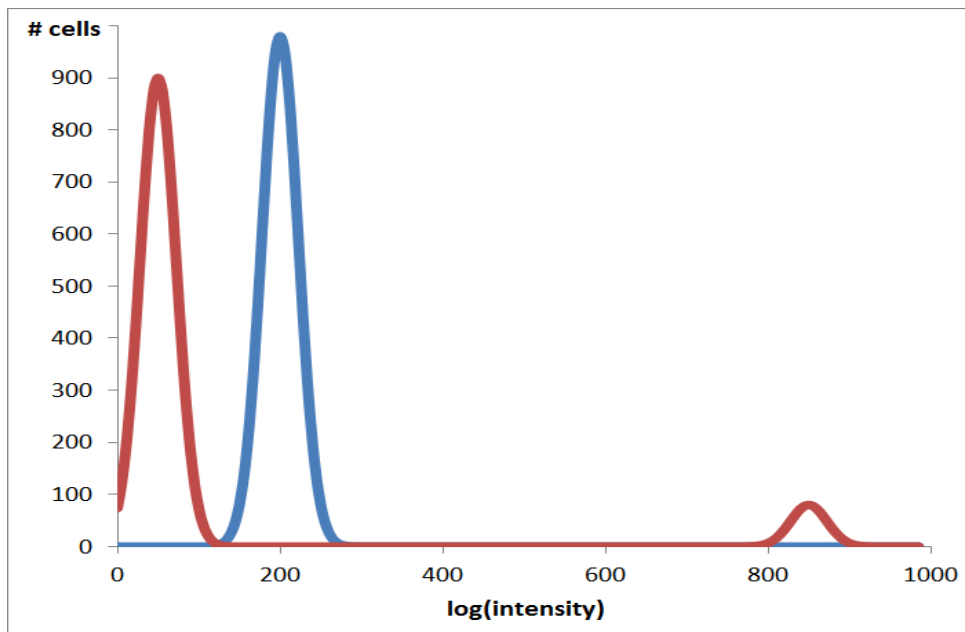
- Light scatter
- Fluorescence
- Phosphorescence
- Raman
- Element mass
- Electrical properties  
e.g. impedance
- ...



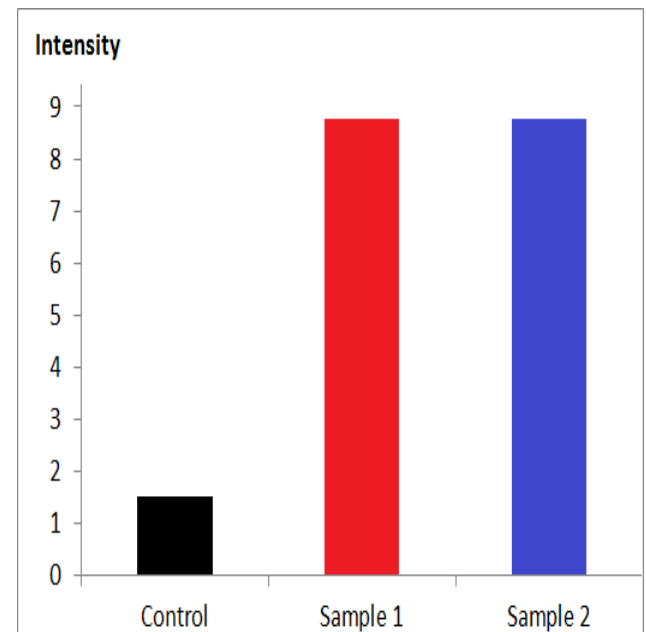
<http://www.dvssciences.com/technical.html>

# Why Single Cell/Particle Analysis

Intensity Histogram for Single Particles



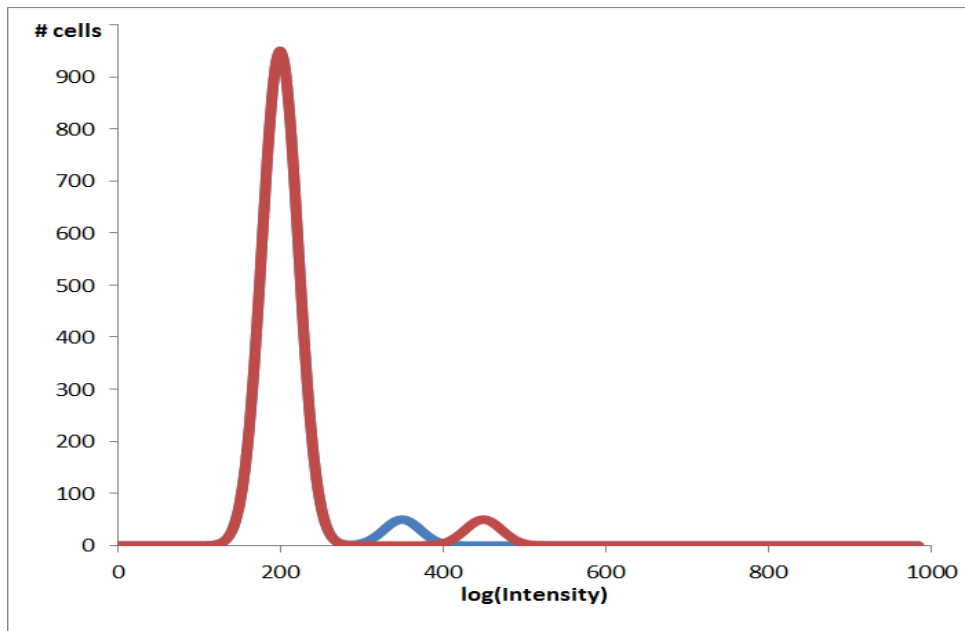
Intensity per Sample



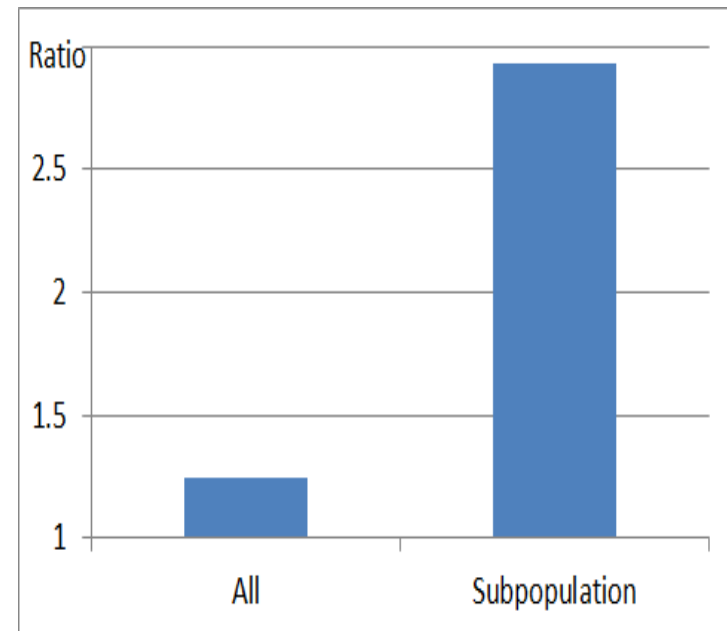
Cell by cell intensity analysis detects population heterogeneity.

# Benefits of Subset Specific Analysis

**Intensity Histogram**



**Intensity Ratios**

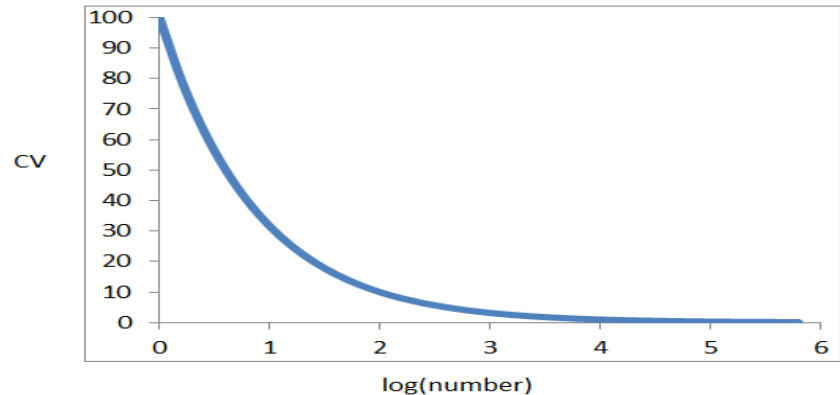
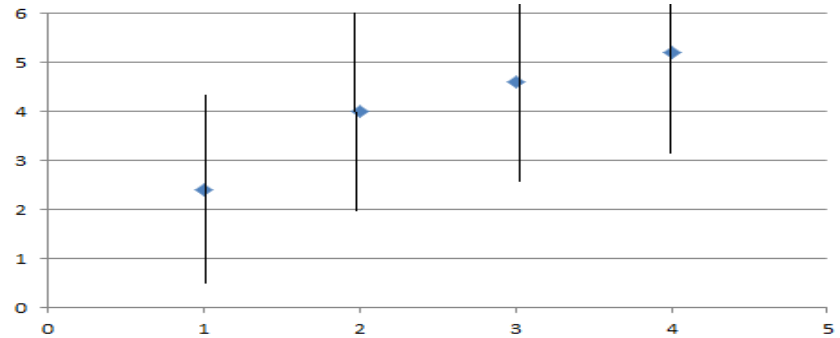


Subpopulation analysis detects changes better, especially for rare subpopulations.

# Cell Counting (abs. counts or percentages)

## Counting Statistics

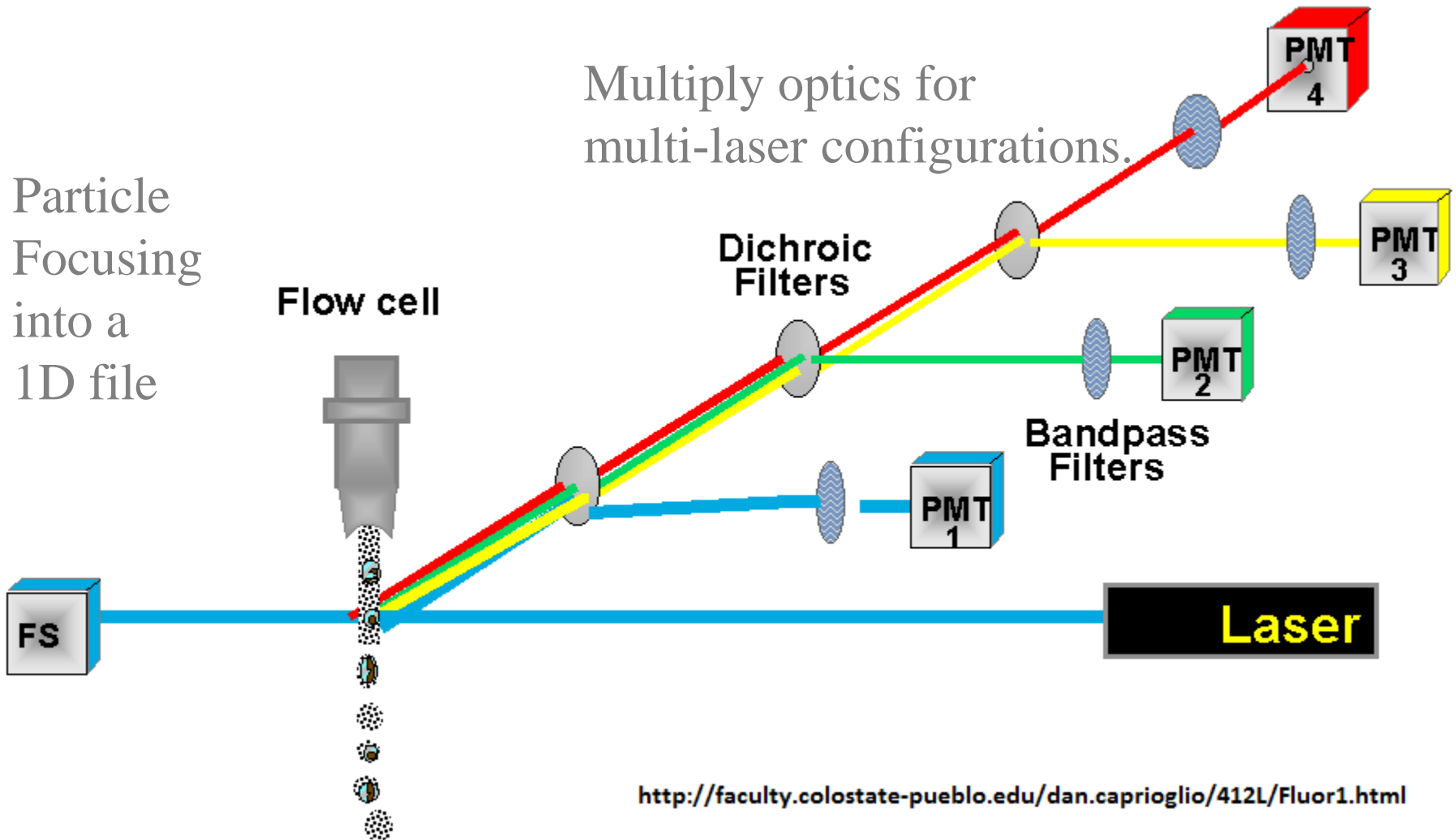
	Sample 1	Sample 2	Sample 3	Sample 4
	6	2	6	8
	3	7	1	6
	1	3	5	3
	1	4	5	6
	1	4	6	3
<b>Mean</b>	2.4	4	4.6	5.2
<b>St.Dev</b>	2.2	1.9	2.1	2.2
		<b>Overall</b>	<b>Mean</b>	4.1
			<b>St.Dev</b>	2.2



Ignoring Counting Statistics Can Lead to  
Erroneous Conclusions

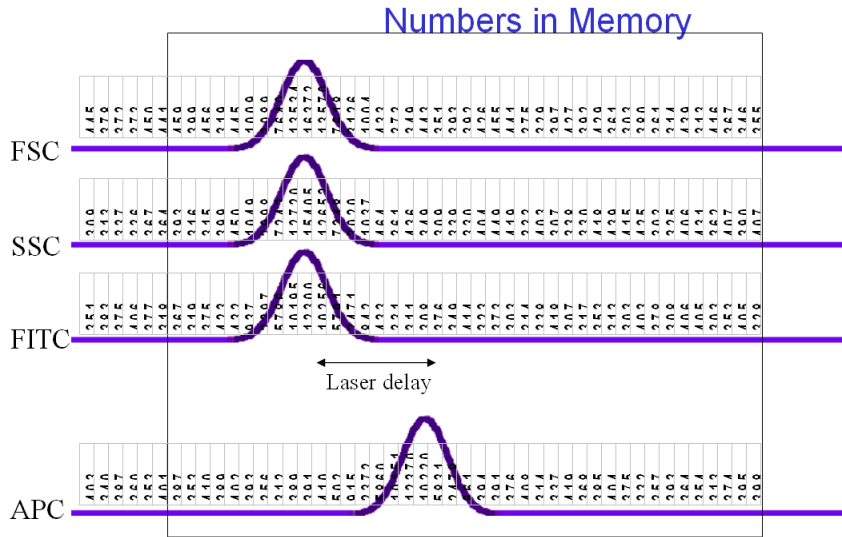


# Flow Cytometer Components

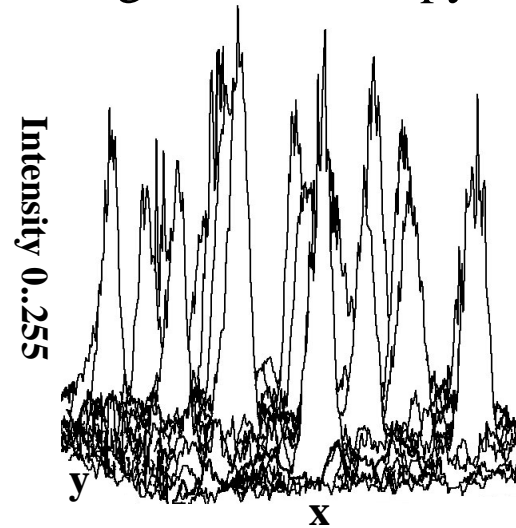


# Basic Data Processing

## Flow Cytometry

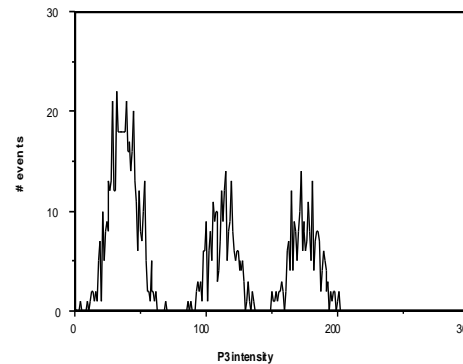


## Digital microscopy

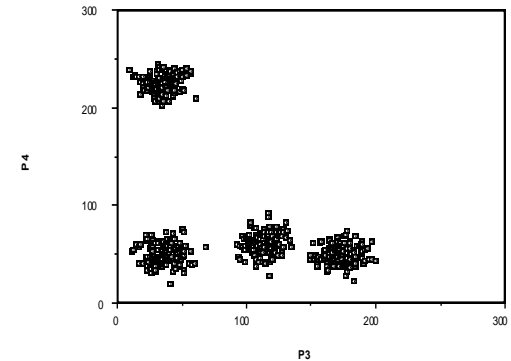


Cell	P1	P2	P3	P4	P5	Pop#
1	242	135	704	175	612	1
2	146	132	690	178	566	1
3	269	147	89	206	580	3
4	442	143	399	250	255	4
5	212	167	155	926	526	2
6	269	2	659	207	575	1
7	204	232	112	171	679	3
8	152	74	160	828	532	2
			...			
9997	215	119	138	936	662	2
9998	244	50	72	261	543	3
9999	214	137	174	1014	597	2
10000	312	87	110	904	560	2

Event histogram



"Dotplot"

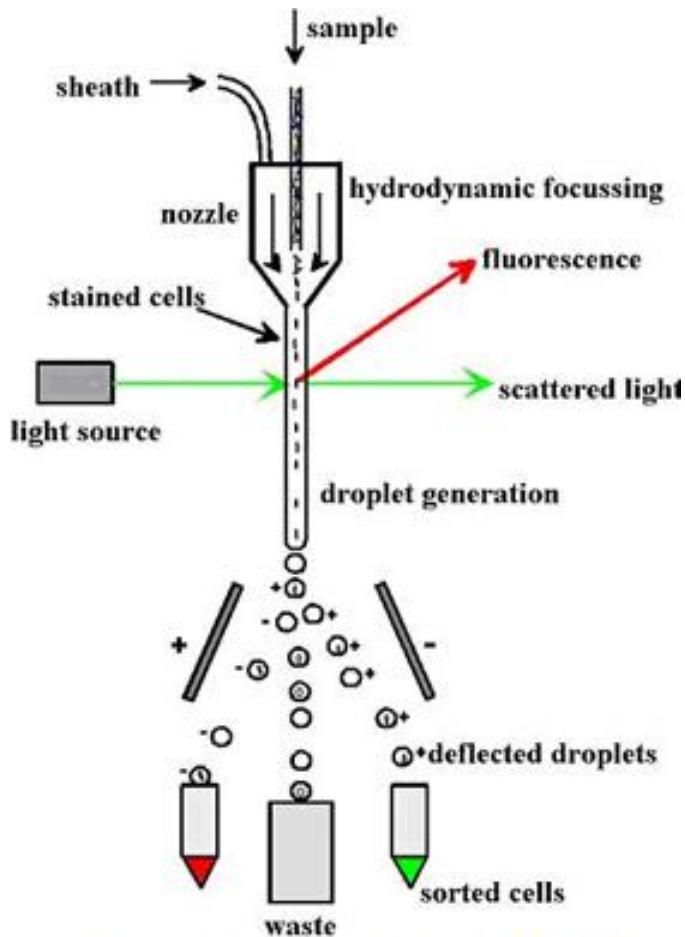


for >2 parameters: gating, cluster analysis, ...  
 For many samples and parameters: bioinformatics

# Cell Sorting

## Applications Examples

- Chromosomes
- Strain Improvement
- Genomics
- Proteomics
- ...



[www.lifesciencesfoundation.org/events-The\\_FACS.html](http://www.lifesciencesfoundation.org/events-The_FACS.html)

Cell sorting review: Derek Davies

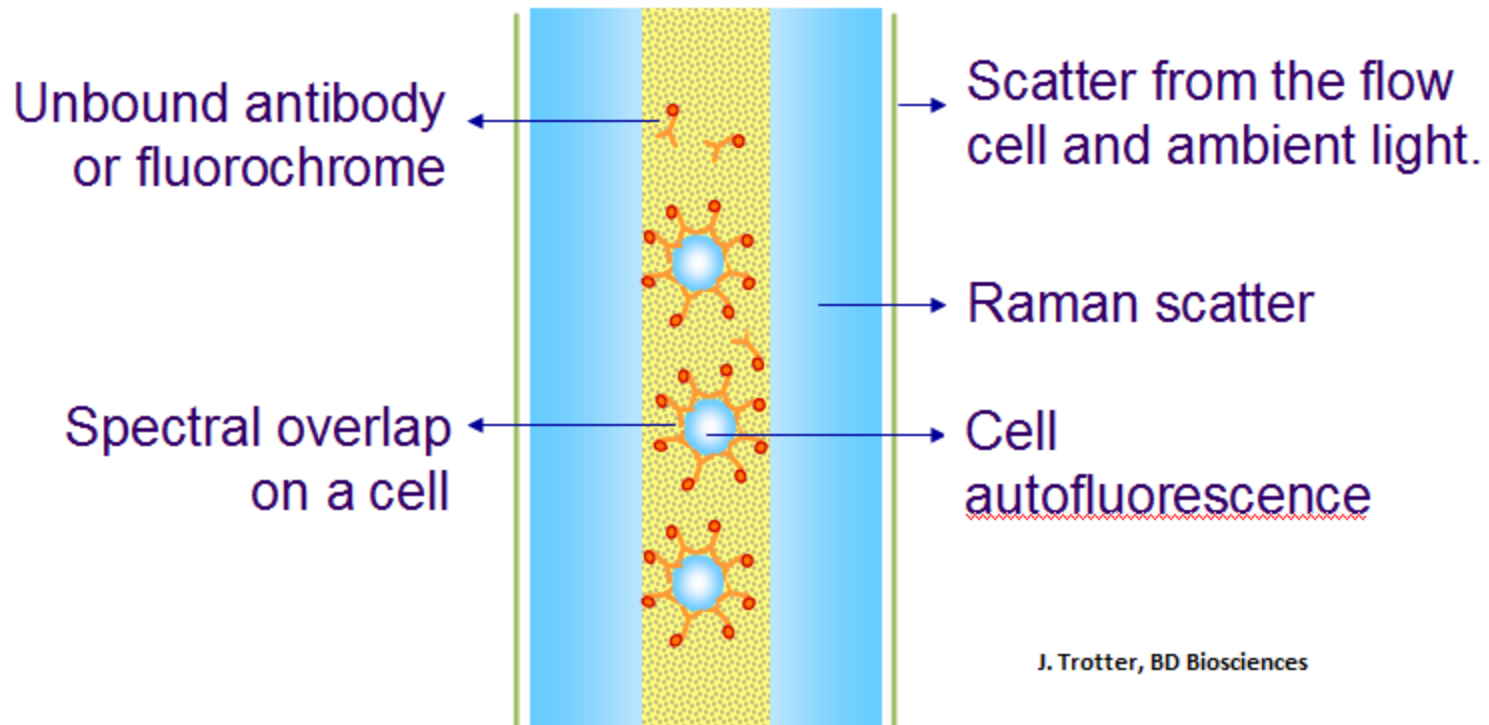
<http://www.facs.ethz.ch/docs/lit>

see also:

<http://www.desatoya.com/ScienceTechnology/CytometryWithSorting.htm>

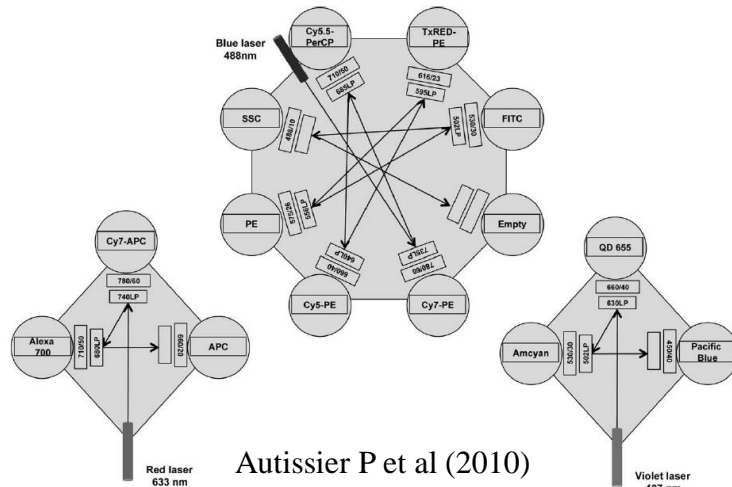
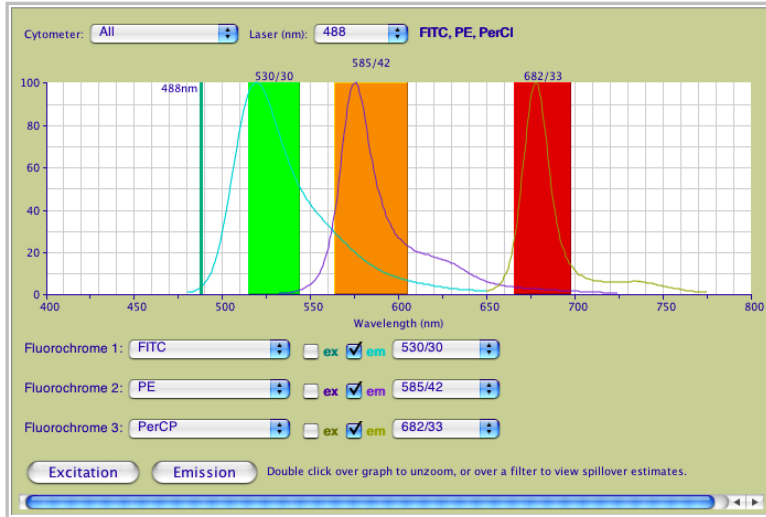
# Instrument Evaluation Br

Relative B (Br) is a measure of true optical background in the fluorescence detector.

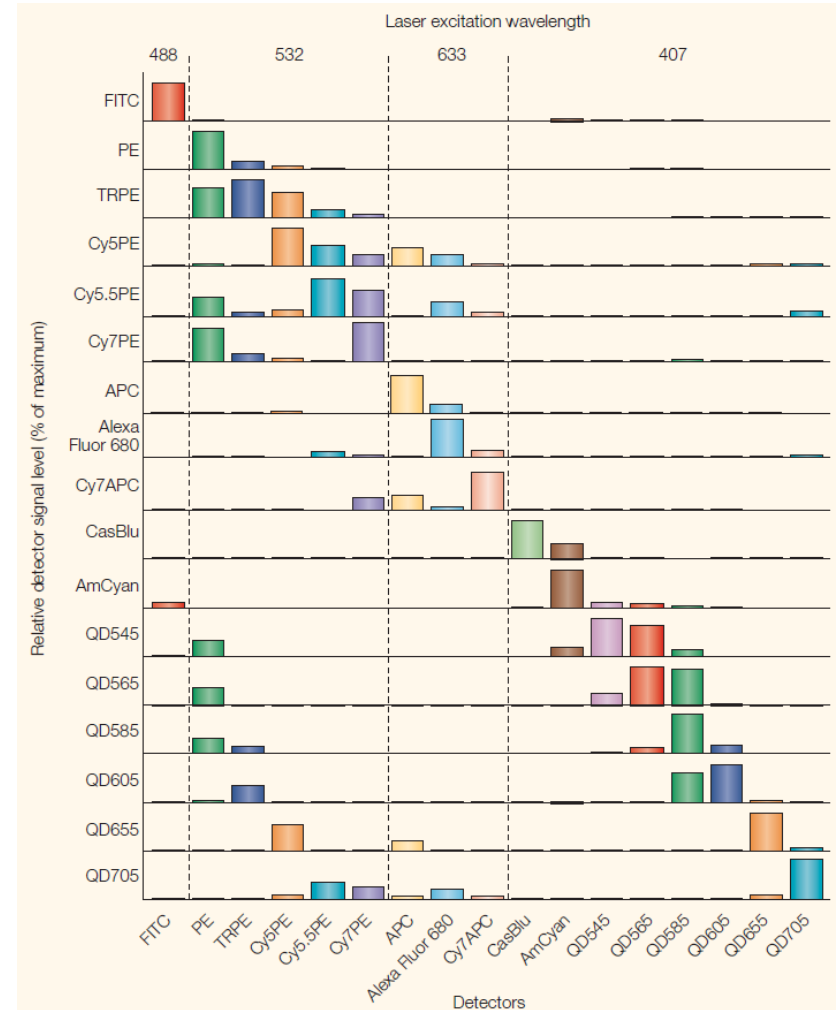


# Filter Arrangement and Spectral Overlap

(not relevant for element mass cytometry)

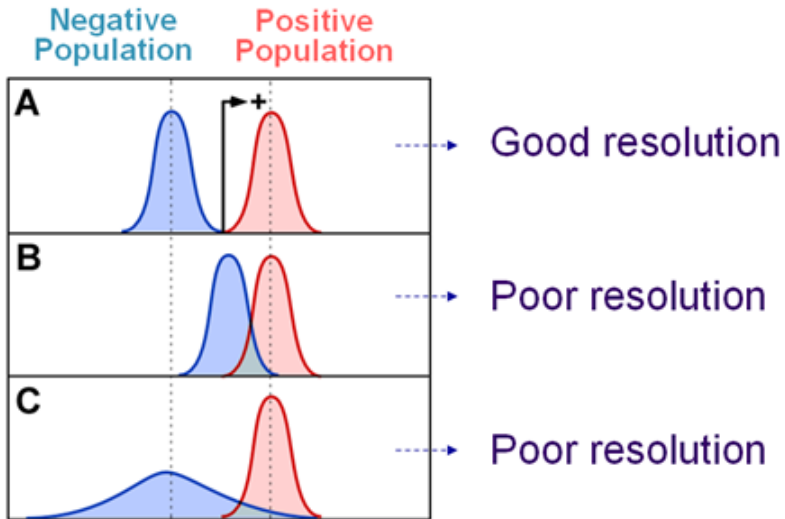


Autissier P et al (2010)  
Cytometry 77A, 410ff

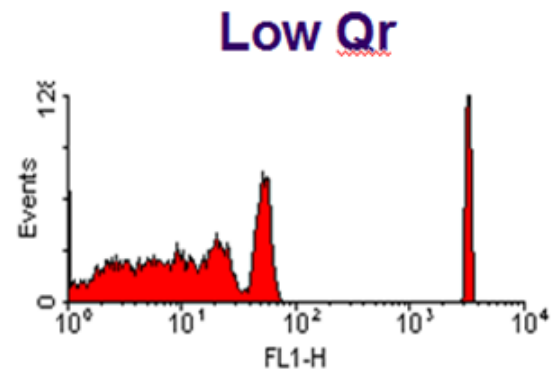
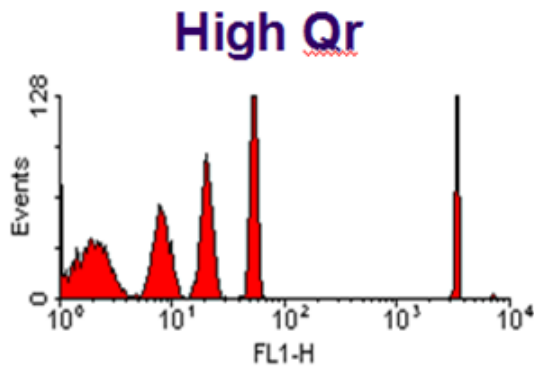


Perfetto SP et al (2004)  
Nature Reviews Immunology 4, 648ff

# Instrument Evaluation Qr



$$Q_r = \frac{\# \text{ photoelectrons}}{\# \text{ fluorescence molecules}}$$



# Optimizing cytometry measurements (I)

- Background light

- The total measurement SD is the sum of the error contributions from all sources:

$$SD = \sqrt{SD_{optical}^2 + SD_{intrinsic}^2 + SD_{illumination}^2 + SD_{noise}^2}$$

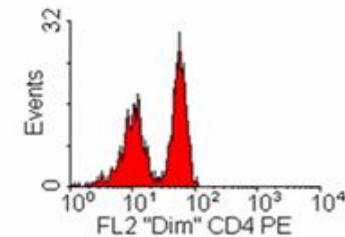
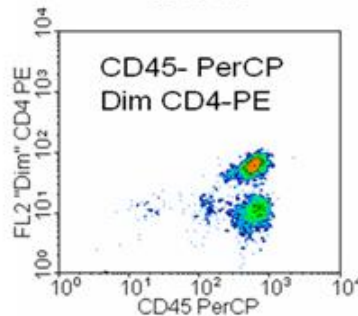
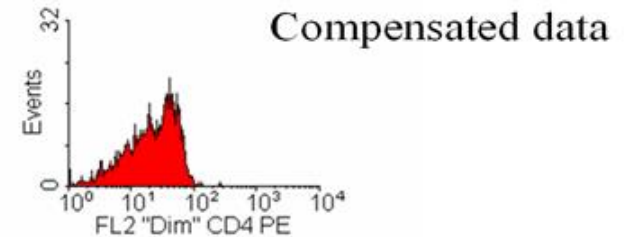
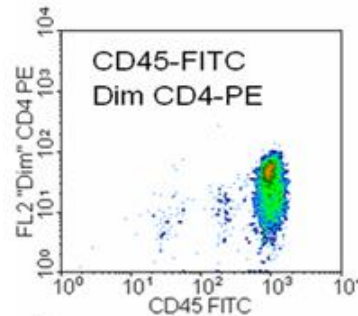
- When two dyes (PE and FITC) are measured by a single detector the SD is additive:

$$SD_{optical} = \sqrt{SD_{PE}^2 + SD_{FITC}^2 + SD_{background}^2}$$

Reagent performance

$$\text{Stain index} = \frac{Medium_{pos} - Medium_{neg}}{2 * SD_{neg}}$$

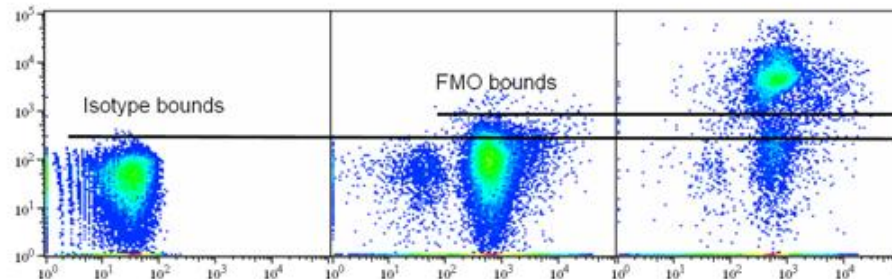
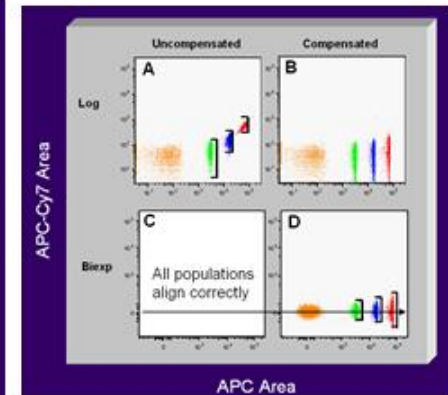
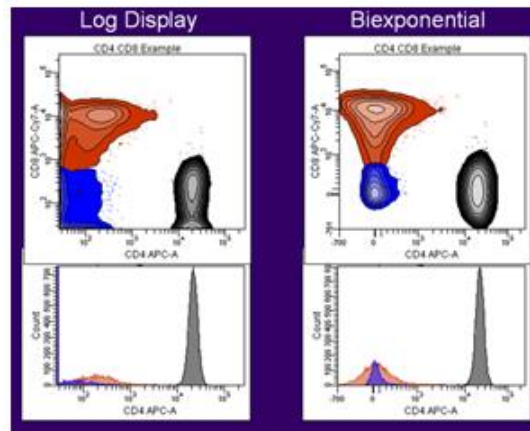
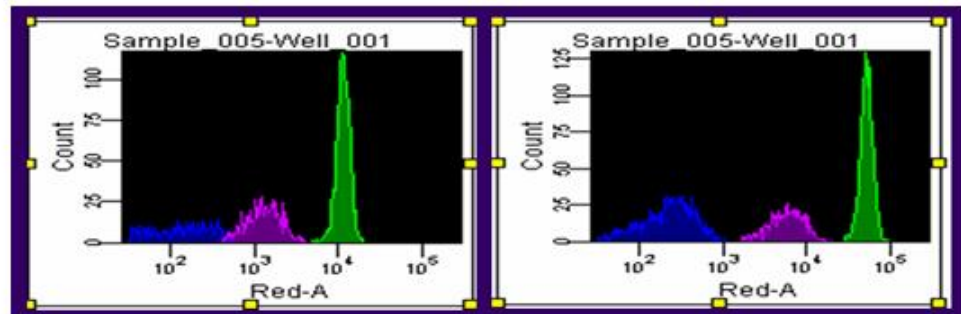
- Dye properties (brightness and spectral overlap)



Better separation with less spectral overlap.

# Optimizing cytometry measurements (II)

- Gain (PMT, CMOS, CCD) settings
- Data Display
- Controls





# Multi-parameter Fluorescence Cytometry

## Points To Consider

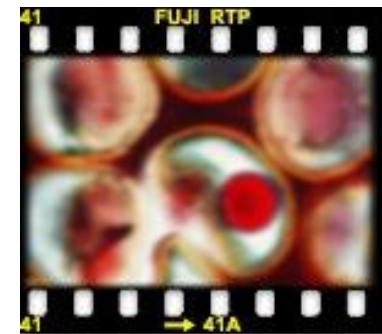
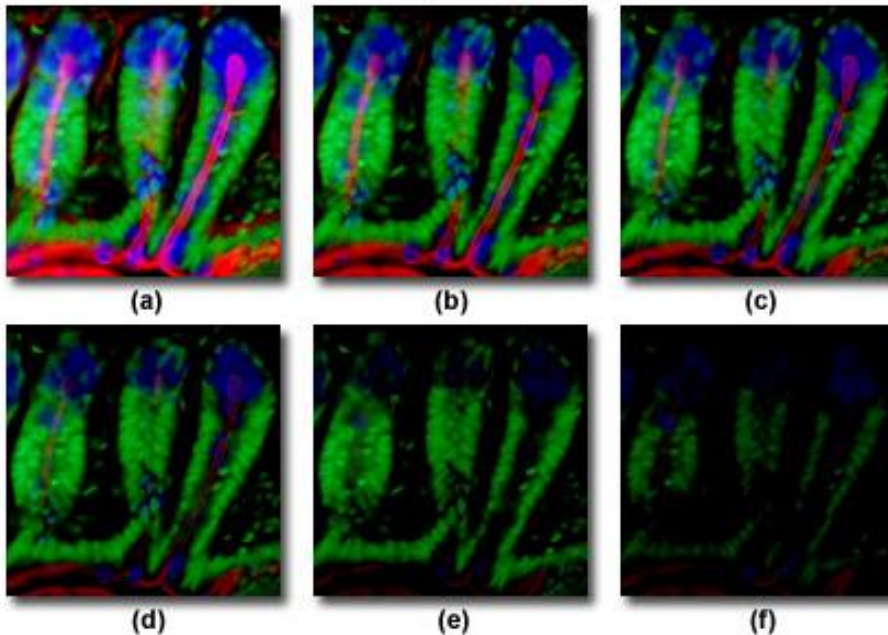
- Know your instrument status e.g. Qr & Br for different channels
- Use high enough gain settings to maximize sensitivity
- An antibody/dye combination that marginally allows discrimination of positives/negatives in a single color assay is unlikely to contribute anything helpful in a multicolor experiment.
- Avoid spillover from bright cell populations into channels requiring high sensitivity
- Beware of tandem dye degradation
- Internal controls are essential

# Quantitative Multi-color Microscopy

## Additional factors

- Field to field focus
- Photobleaching

Differential Photobleaching in Multiply-Stained Tissues



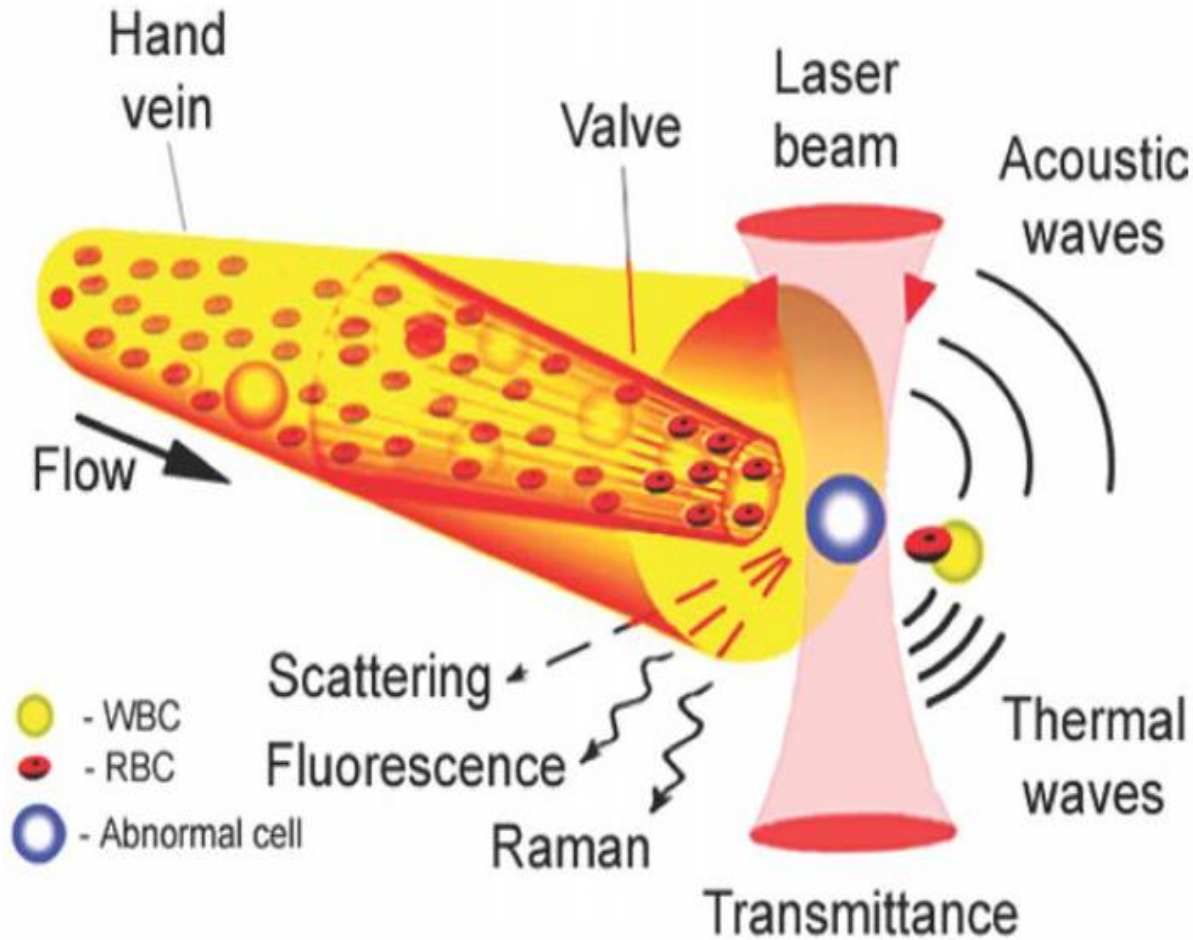
Out of Focus

Images from

<http://micro.magnet.fsu.edu/primer/index.html>

# In-vivo Multi-parameter Cytometry

Single cell analysis in living animals

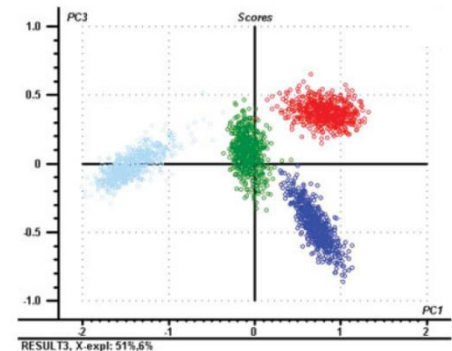
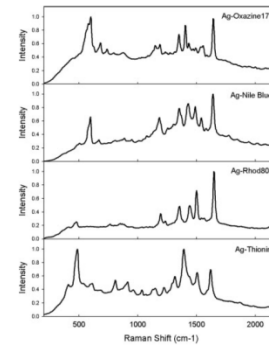
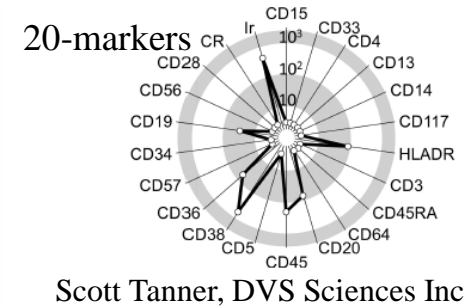
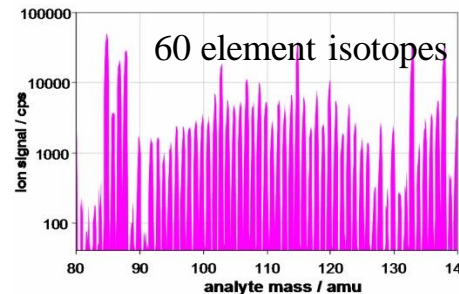


Issues:

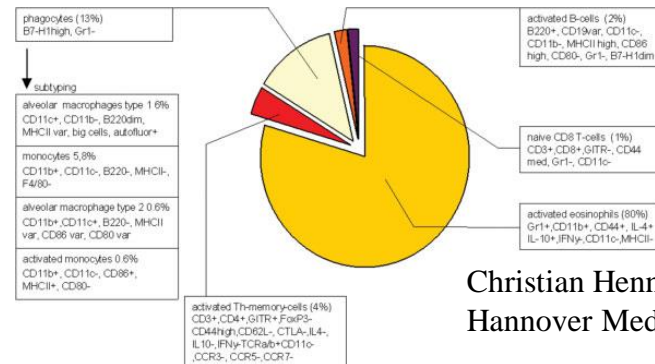
- tissue optics
- object motion
- flow rate
- labelling
- ...

# New Developments for in-vitro Multi-parameter Cytometry

- **Element-Label Flow Cytometry (CyTOF, addresses fluorescence spectral overlap issue by using elements as labels, Anal. Chem., 2009, 81 (16), pp 6813–6822)**
- **SERS-Label and Spectral Flow Cytometry (uses spectral (fine)-structure to distinguish labels, Cytometry, 2008, 73A(2), pp 119-128, SONY cytometer)**
- **Sequential Stain De-stain Cytometry (Cytometry, 2009, 75A(4), pp 362-370)**



John Nolan, La Jolla Bioengineering Institute



31-marker analysis

Christian Hennig, ChipCytometry Hannover Medical School

# Conclusions

## Multi-parameter cytometry

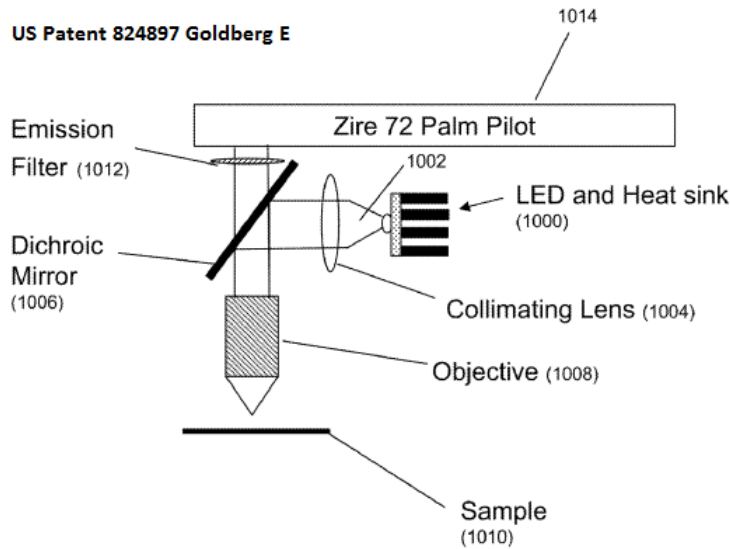
Optimized flow and imaging single cell cytometry with adequate bio-informatics tools provide quantitative molecular measurements into biological processes at organism, cellular and sub-cellular levels.

New developments in many areas have simplified the tools for the biologist.

# Evolving Technologies for Cytometry

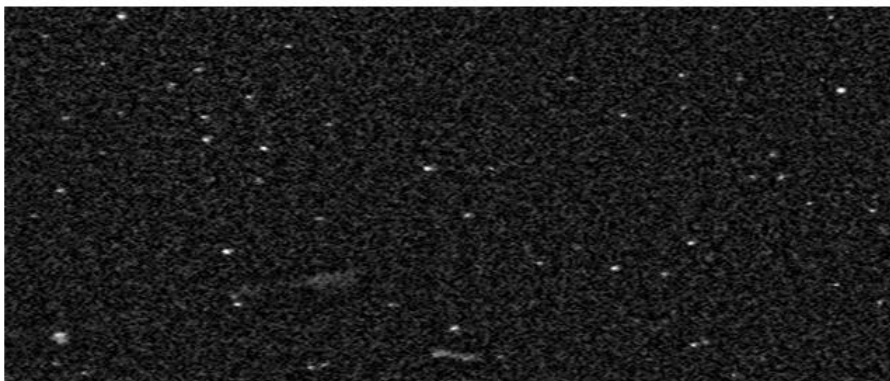
- More low complexity cytometers for cell (subset) counting
- Novel cell sorters
- Integrated Cell Analysis System
- Intra-vital imaging
- Innovative sample preparation
- Fluorescent polymers for high sensitivity
- Novel affinity reagents (antibodies)

# Low-complexity Cytometers for Cell Counting

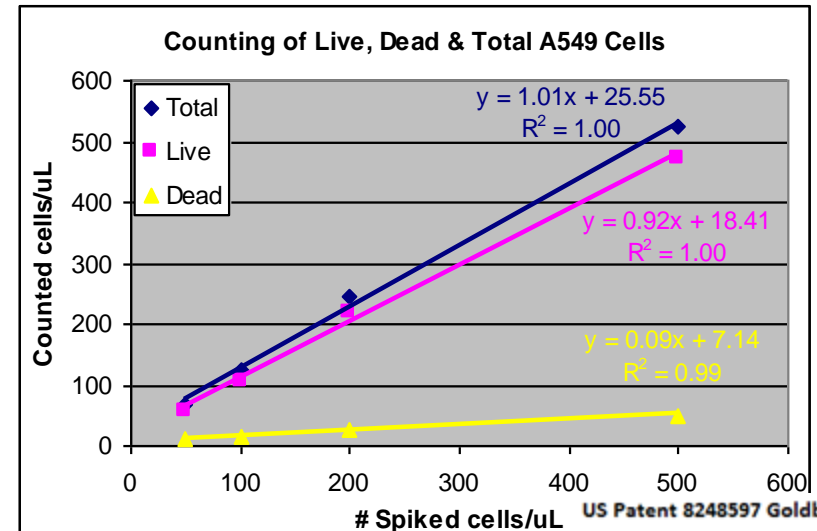


## Counting Nucleated Cells

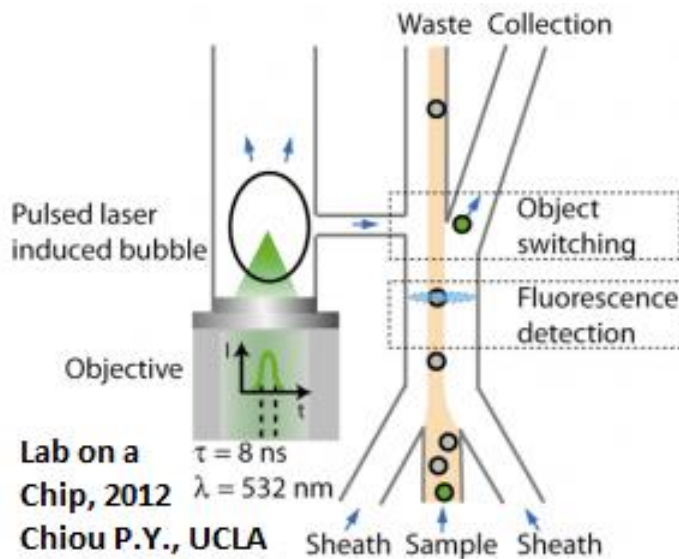
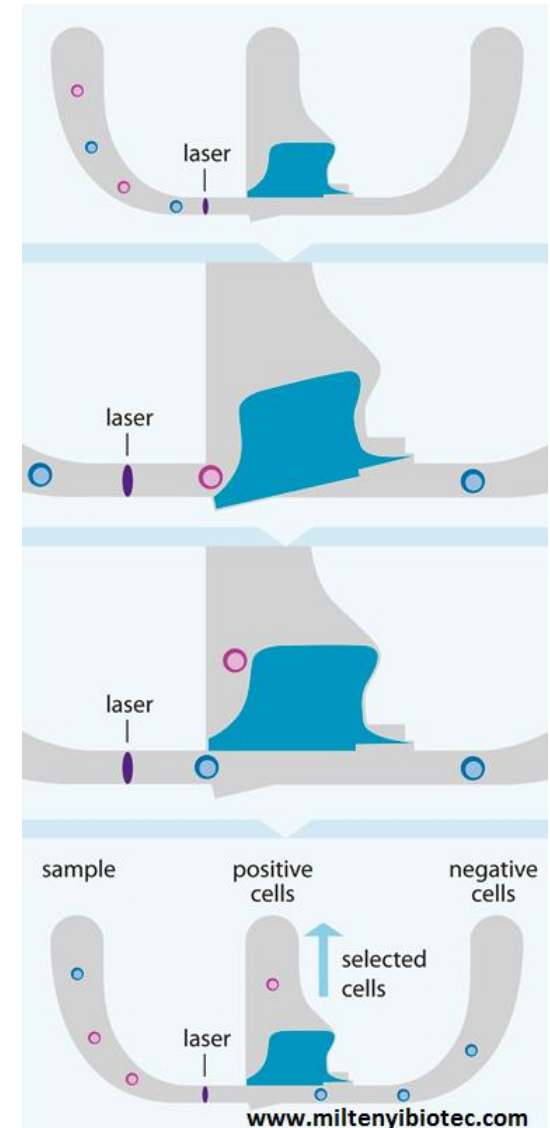
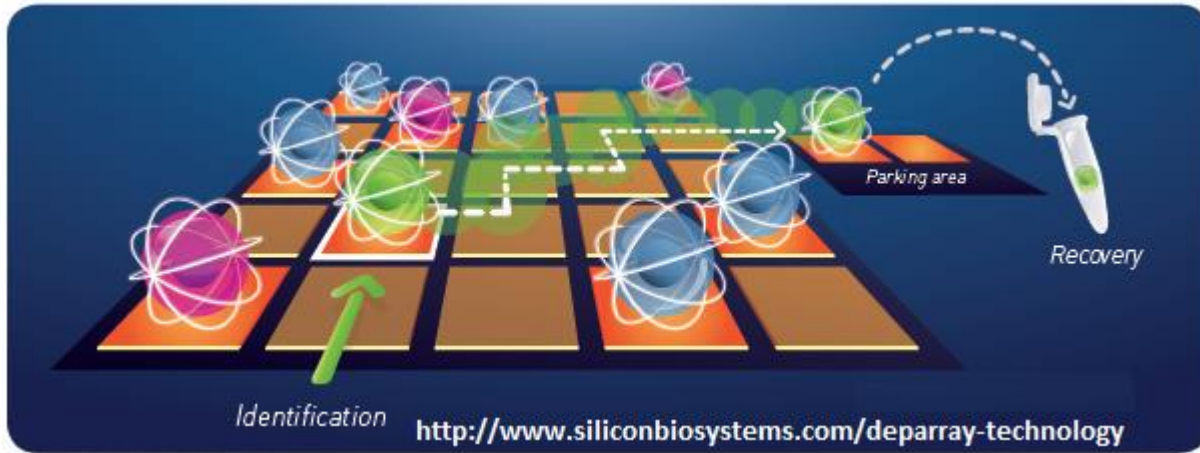
Propidium Iodide (impermeant) + SYTO-17 (permeant)



US Patent 8248597 Goldberg E

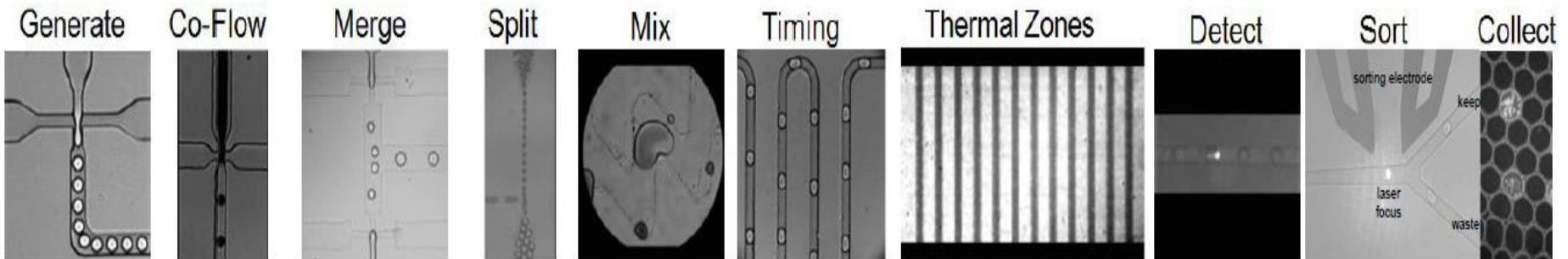
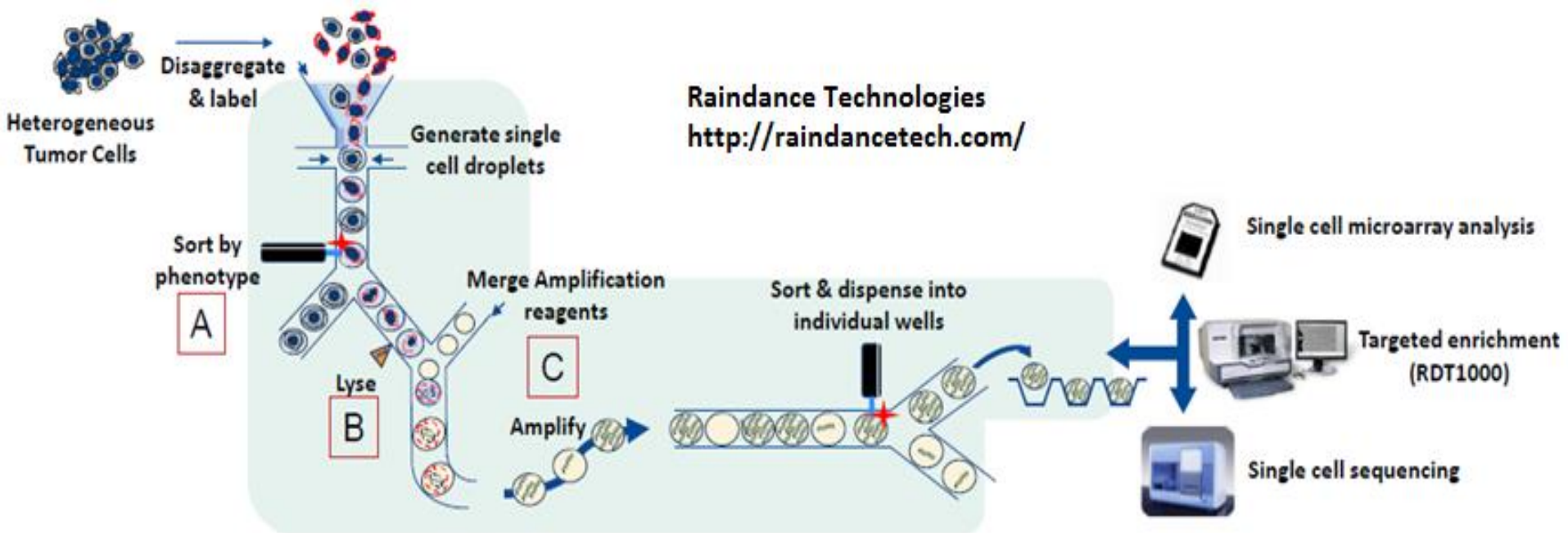


# Novel Cell Sorters



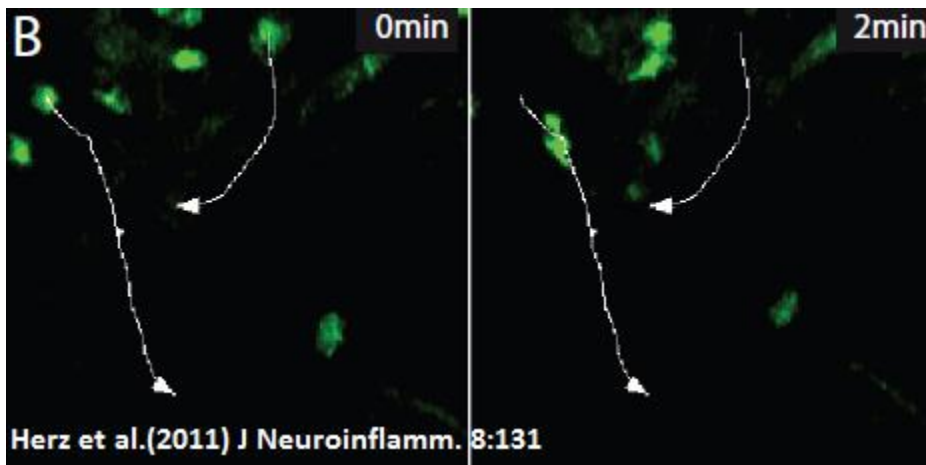
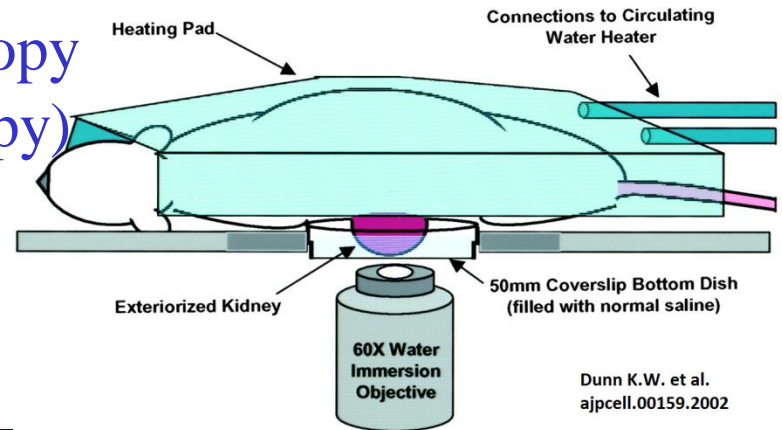


# Integrated Bio-Assay System



# Intra-vital Imaging

- Two-photon laser scanning microscopy
- Raman (SERS and CARS microscopy)
- Positron emission tomography
- Ultrasound, x-Rays
- ...



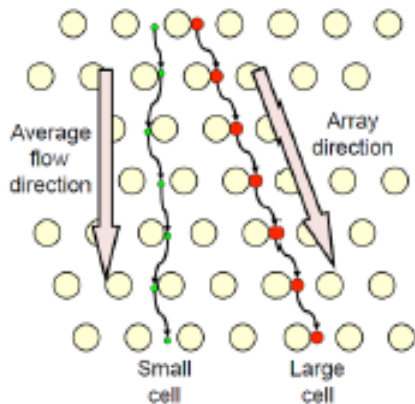
## Issues:

- tissue optics
- object motion
- flow rate
- labeling
- ...

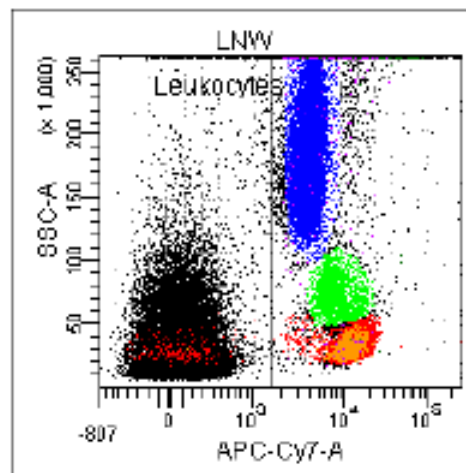
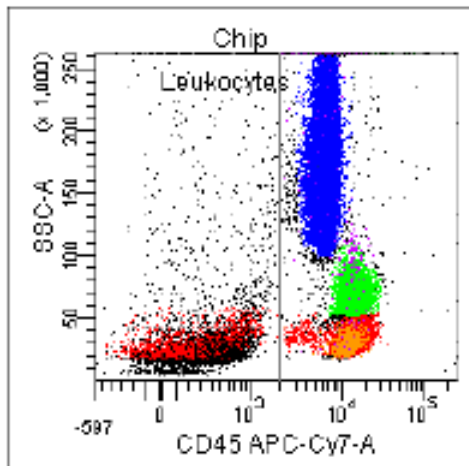
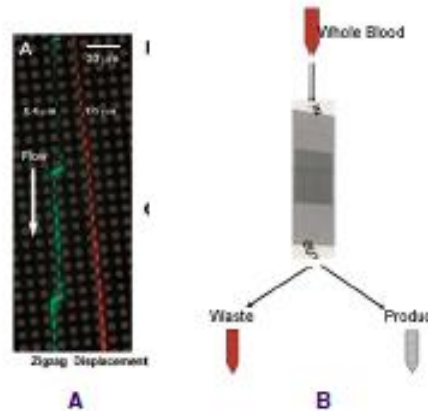
**Recent review of in-vivo microscopy:** Andresen V, et al. (2012) High-Resolution Intravital Microscopy. *PLoS ONE* 7(12): e50915

# Innovative Sample Preparation

Microfluidic system for leukocyte isolation and cell washing  
(deterministic lateral displacement)



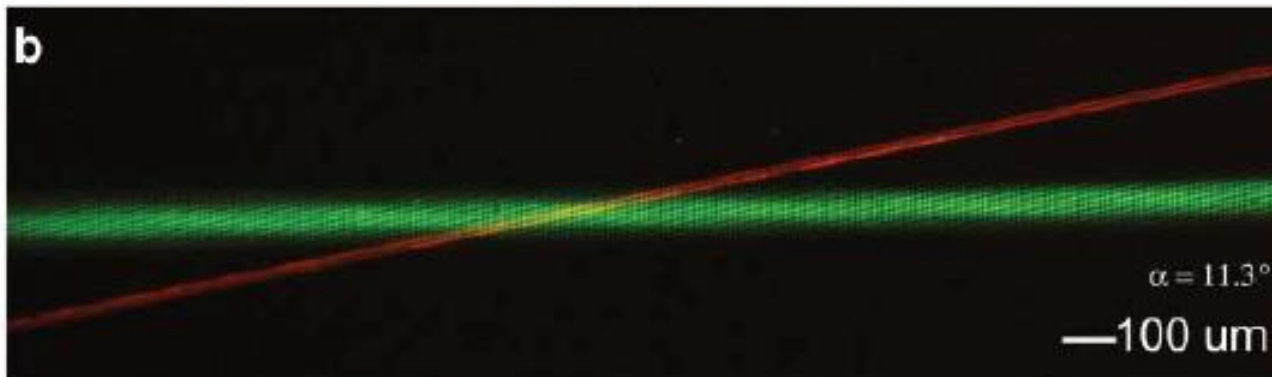
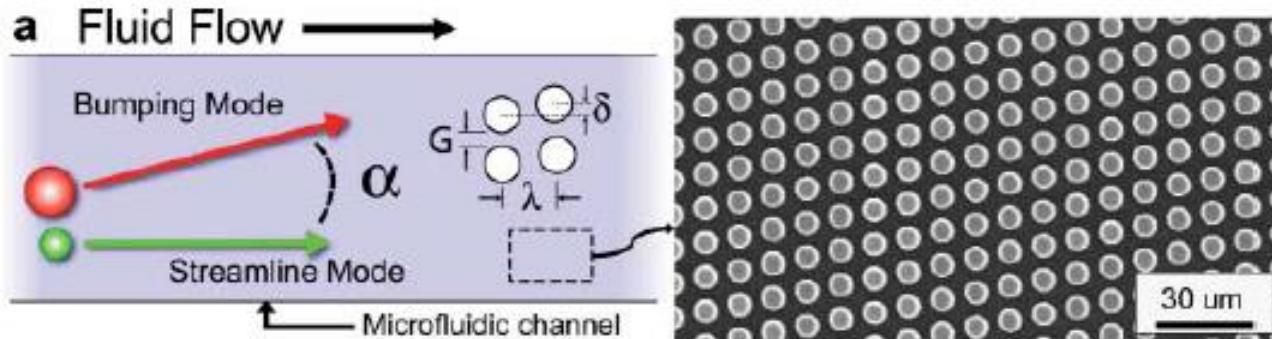
Chip and new blood separation process



also:

- acoustic focusing
- microfluidic filters
- inertial flow
- magnetic nanoparticles
- high density particles
- dielectrophoresis
- optical traps
- ...

# Deterministic Lateral Displacement



Separating particles by size, adding reagent, mixing, and washing

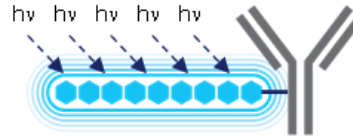
From: Morton K J et al.

Lab on a Chip 2008

# Bright Fluorescent Polymer Dyes (BD Sirigen)

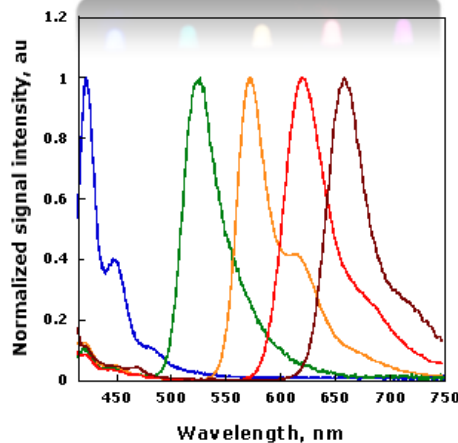
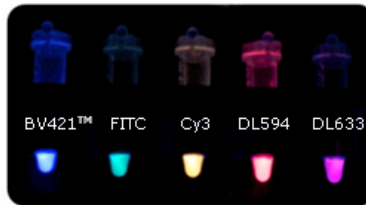
## Polymer Based Fluorochromes

- Well defined synthetic organic polymer structures
  - Single conjugation site, defined size, etc.
- Backbone comprised of  $\pi$ -conjugated repeat units
  - Affords massive light harvesting ( $\epsilon > 10^6$ ) materials with high quantum yields
- Tunable architecture adapted for low NSB, high aqueous solubility and spectral performance



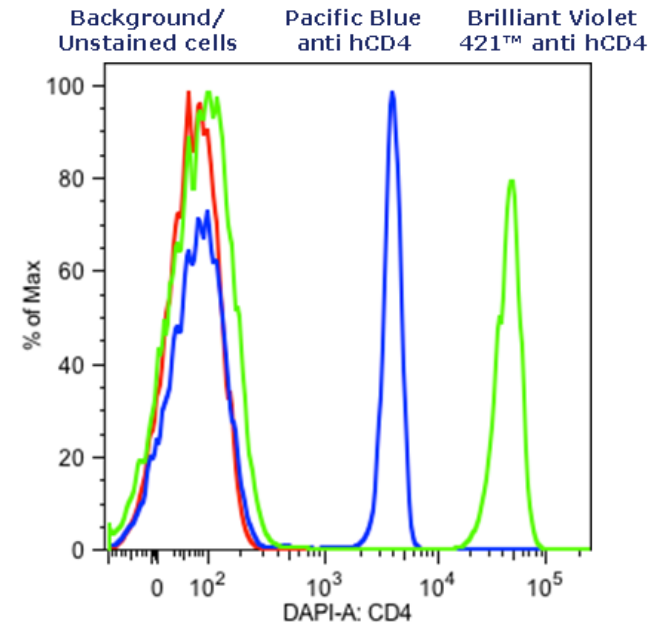
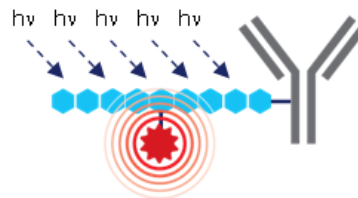
## Brilliant Violet 421™

- PE level performance w/ 405nm Laser
- >10x the Stain Index of Pacific Blue
- Enables detection of low abundance targets in multicolor assay panels (e.g. CD56, CD127, etc.)
- Wide range of Ab clones validated



## Brilliant Violet Tandems

- Provides a wider range of colors spanning the visible spectrum
  - >6 unique colors validated
- Chemically controlled ratio of donor/acceptor provides:
  - Reproducible performance
  - Low (<5%) compensation at 450nm



see also: Cytometry A. **81**: 45ff

[http://sirigen.com/flow\\_cytometry.html](http://sirigen.com/flow_cytometry.html)

# Novel Affinity Reagents

- Antibodies
  - Antibodies from different species (e.g. Llama 15 kDalton fragments with  $10^{-9}$ M Kd and high stability, potential for intracellular use)
  - Recombinant antibody fragments
  - ...
- Synthetic affinity reagents
  - Aptamers
  - Protein scaffolds
  - Molecular Imprinted Polymers

# Conclusions

## Evolving Technologies

Technology developments in algorithms, computing, detectors, electronics, nanotechnology, microfluidics, organic chemistry, and recombinant protein technology create the basis for new reliable analytical approaches for a deeper molecular understanding of living systems.

There is value in working with other scientific disciplines.

# Acknowledgements

- Joe Trotter
- Ming Yan
- Maria Jaimes
- Brian Warner
- Ed Goldberg
- Hrair Kirakossian
- Liping Yu
- Brent Gaylord
- Mike Brasch
- Ben Verwer
- ...  
above all BD
- Holden Maecker, Stanford
- Bob Hoffman, consultant
- Martin Buescher, Miltenyi
- ...
- BD Biosciences
- AmCell Corp/ Miltenyi Biotec
- ...

## Contact

**Email:** [diether@desatoya.com](mailto:diether@desatoya.com)

**Phone:** USA-408-658-6074

**URL:** <http://www.desatoya.com>